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Short communication

Crimean-Congo hemorrhagic fever serosurvey in at-risk professionals, Madagascar, 2008 and 2009

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ABSTRACT

Background: Crimean-Congo hemorrhagic fever (CCHF) is a zoonotic arboviral infection with hemorrhagic manifestation and often a fatal ending. Human become infected mainly through tick bite or by crushing infected tick, by contact with blood or tissues from viraemic livestock or patient. CCHF virus (CCHFV) has been isolated once in Madagascar but data on the epidemiology of the disease in the country are very scarce.

Objectives: To investigate the circulation and the geographic distribution of CCHFV infection among at-risk population in Madagascar.

Study design: A national cross-sectional serologic survey was performed in 2008–2009 among slaughterhouse workers.

Results: A total of 1995 workers were included. A recent CCHFV infection was detected in 1 of the 1995 participants (0.5%; 95% confidence interval [CI]: 0–0.15%), and a past CCHFV infection was detected in 15 participants (0.75%; 95% CI: 0.37–1.13%).

Conclusion: Overall, the percentage of CCHFV infection seen in Madagascar among at-risk professionals is very low compared to endemic countries. An assessment of the prevalence in livestock as a sensitive indicator of CCHFV activity must be considered in order to confirm the lack or the weak endemicity of CCHF in Madagascar.

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1. Background

Crimean-Congo hemorrhagic fever (CCHF) is a tick-borne disease caused by a virus (CCHFV) belonging to the family Bunyaviridae, genus Nairovirus (reviewed in 1, 2). This zoonosis is largely distributed in Africa, Asia, Middle East and southern Europe (Balkan Peninsula). Hyalomma spp. is the most important and widely distributed tick vector, but other genera (Rhipicephalus, Boophilus, Dermacentor, and Ixodes) may have been contributed to CCHFV ecological cycle. Ruminants, but also small terrestrial mammals and birds are involved in an enzootic tick-vertebrate-tick cycle. Human become infected through tick bite or by crushing infected tick, by contact with blood or tissues from viraemic livestock, or by unprotected contact with biological fluids of a CCHF patient during the acute phase of infection. Disease is seen only in humans and frequently fatal with severe hemorrhagic signs. Treatment is symptomatic and to date, no vaccine is available.1,2

In Madagascar, CCHFV was isolated only once, from Rhipicephalus (Boophilus) microplus ticks collected on cattle, in March 1985, in the main slaughterhouse in Antananarivo. Animals were coming from Tsiranoamandidy, in the highlands, 150 km West from Antananarivo. It was the largest live market in the country receiving cattle from all places in Madagascar.3

Phylogenetic studies based on partial S sequences indicated that the Malagasy strain was closer to strains isolated in Middle-East and Asia than to African isolates.4 The only serological evidence of CCHFV human infection (using immunofluorescence assay) was demonstrated in 2 out of 149 individuals sampled in 1988 in Mandoto, a cattle breeding area in the highlands.5

Abbreviations: CCHF, Crimean-Congo hemorrhagic fever; CCHFV, Crimean-Congo hemorrhagic fever virus; ELISA, enzyme-linked immunosorbent assay; CDC, Centers for Disease Control and Prevention.

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Data on CCHF epidemiology in Madagascar are consequently very limited.

2. Objectives

To investigate the circulation and the geographical distribution of CCHFV infection among at-risk population in Madagascar.

3. Study design

We performed a nationwide serological survey, from September 2008 through May 2009, among 1995 human volunteers at high risks of zoonotic infection as CCHF.6 These volunteers were people living in 106 out of 111 administrative districts of Madagascar and working in slaughterhouses, exposed to fresh meat or blood of livestock since at least 2007. The study was approved by the Malagasy National Ethical Committee. Informed written consent was obtained from the participants. Detection of IgM and IgG antibodies against CCHFV was performed by ELISA as previously described.7 Briefly, following heat and detergent inactivation, sera were tested by CCHFV-specific IgM and IgG ELISAs. The assays were completed using inactivated CCHFV-infected Vero E6 cell antigens and uninfected Vero E6 cell antigens, and using four dilutions of each serum (1/100, 1/400, 1/1600, 1/6400). Titers and the cumulative sum of optical densities of each dilution (SUMOP) minus the background absorbance of uninfected control Vero E6 cells (adjusted SUMOP) were recorded. Results of the assays for sera were considered positive only if the adjusted SUMOP and titer were above preestablished conservative cutoff values, which were set for IgM ELISA (≥0.75 and ≥1/400) and IgG ELISA (≥0.95 and ≥1/400). Positive samples and 3% of the negative samples tested in the Institut Pasteur in Madagascar (IPM) were sent to the Institut Pasteur in Lyon and to the CDC in Atlanta to validate the IPM ELISA results.

4. Results

A total of 1995 persons, aged 15–85 years participate to the study. The median age was 34 years (36 missing data). The sex ratio was 13.7 (six missing data). A recent CCHFV infection (presence of IgM against CCHFV and lack of IgG against CCHFV) was detected in 1 of the 1995 participants (0.5%; 95% confidence interval [CI]: 0–6.15%), and a past CCHFV infection (presence of IgG against CCHFV and lack of IgM against CCHFV) was detected in 15 participants (0.75%; 95% CI: 0.37–1.13%). Titers were 400, 1600, and 6400 for 11, 3 and 1 participants, respectively. CCHFV antibody positive subjects were detected in 14 of the 106 districts tested suggesting a scattered distribution (Fig. 1). The seropositivity was not significantly found associated with age, sex or location of activity of participants (data not shown).

5. Discussion

Overall, the percentage of CCHFV infection seen in Madagascar among at-risk professionals is very low compared to those observed in endemic countries like Mauritania (7%) and United Arab Emirates (6%).7,8 This observation may be explained by the lack of ticks of the genera Hyalomma in Madagascar,9 Rhipecephalus (Boophilus) microplus, the species found infected by CCHFV in Madagascar is widely distributed in the country up to 1950 m of altitude (Stachurski, pers. comm.). However, the vector competence of this species has not been demonstrated in the laboratory.2

The low percentage of detection of human antibodies against CCHFV and the scattered geographic distribution may be the consequence of repeated introductions of infected animals, large movements of domestic ruminants in the country, and abortive circulations of CCHFV within the country. For Rift Valley fever, we have genetic evidence that outbreaks in Madagascar resulted from multiple virus introductions from the east Africa mainland rather than enzootic maintenance.10 Livestock movements were already implicated in the large diffusion of RVFV during the 2008–2009 outbreak.6

Since the first investigations carried out by the Institut Pasteur in the 1970s, 16 arboviruses or related viruses have been isolated in Madagascar,9,11–13 CCHFV is the only member of the genus Nairovirus detected in the island. However, we cannot exclude the presence of an undetected nairovirus close to CCHFV like viruses from the Nairobi sheep disease group including the eponym virus and Dugbe virus, present and widespread in continental Africa (http://www.cdc.gov/nczved/divisions/dvbid/arbovirus.html). Consequently, the occurrence of cross-reaction in our detection of antibodies against CCHFV, if any, highlights the very low circulation of CCHFV in Madagascar.
An assessment of the prevalence in livestock as a sensitive indicator of CCHFV activity must be considered in order to confirm the lack or the weak endemicity of CCHF in Madagascar.

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Competing interests

There is no conflict of interest and absence of any relationship or any degree of conflicting or dual interest, financial or of any other nature that may affect professional judgment in relation to this article.

Ethical approval

Ethical Approval was given by the “Comité National d’Ethique” of Madagascar. Judgement’s reference number: O22-CE/MINSAN.

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