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A molecular marker of artemisinin-resistant *Plasmodium falciparum* malaria

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***Plasmodium falciparum* resistance to artemisinin derivatives in southeast Asia threatens malaria control and elimination activities worldwide. To monitor the spread of artemisinin resistance, a molecular marker is urgently needed. Here, using whole-genome sequencing of an artemisinin-resistant parasite line from Africa and clinical parasite isolates from Cambodia, we associate mutations in the PF3D7_1343700 kelch propeller domain ('K13-propeller') with artemisinin resistance *in vitro* and *in vivo*. Mutant K13-propeller alleles cluster in Cambodian provinces where resistance is prevalent, and the increasing frequency of a dominant mutant K13-propeller allele correlates with the recent spread of resistance in western Cambodia. Strong correlations between the presence of a mutant allele, *in vitro* parasite survival rates and *in vivo* parasite clearance rates indicate that K13-propeller mutations are important determinants of artemisinin resistance. K13-propeller polymorphism constitutes a useful molecular marker for large-scale surveillance efforts to contain artemisinin resistance in the Greater Mekong Subregion and prevent its global spread.**

The emergence of *Plasmodium falciparum* resistance to artemisinin derivatives (ART) in Cambodia threatens the world's malaria control and elimination efforts^{1,2}. The risk of ART-resistant parasites spreading from western Cambodia to the Greater Mekong Subregion and to Africa, as happened previously with chloroquine- and sulphadoxine/pyrimethamine-resistant parasites^{3–5}, is extremely worrisome. Clinical ART resistance is defined as a reduced parasite clearance rate^{1,6–10}, expressed as an increased parasite clearance half-life^{11,12}, or a persistence of microscopically detectable parasites on the third day of artemisinin-based combination therapy (ACT)². The half-life parameter correlates strongly with results from the *in vitro* ring-stage survival assay (RSA_{0–3h}) and results from the *ex vivo* RSA¹³, which measure the survival rate of young ring-stage parasites to a pharmacologically relevant exposure (700 nM for 6 h) to dihydroartemisinin (DHA)—the major metabolite of all ARTs. However, the present lack of a molecular marker hampers focused containment of ART-resistant parasites in areas where they have been documented and hinders rapid detection of these parasites elsewhere, where ACTs remain the most affordable, effective antimalarials. To detect and monitor the spread of ART resistance, a molecular marker for widespread use is needed.

Recent genome-wide analyses of *P. falciparum* isolates have provided evidence of recent positive selection in geographic areas of ART resistance^{9,14–16}. Whereas parasite heritability of the clinical phenotype

is above 50%, no reliable molecular marker has yet been identified. One possible explanation is that the parasite clearance half-life is not only determined by the intrinsic susceptibility of a parasite isolate to ART, but also by its developmental stage at the time of ART treatment and host-related parameters such as pharmacokinetics and immunity¹⁷. This issue was recently highlighted in patients presenting discordant data between parasite clearance half-life *in vivo* and RSA_{0–3h} survival rate *in vitro*¹³. Moreover, genome-wide association studies (GWAS) are confounded by uncertainties about parasite population structure. Recent evidence for several highly differentiated subpopulations of ART-resistant parasites in western Cambodia¹⁵ suggests that distinct emergence events might be occurring. An alternative strategy to discover a molecular marker is to analyse mutations acquired specifically by laboratory-adapted parasite clones selected to survive high doses of ART *in vitro*, and use this information to guide analysis of polymorphism in clinical parasite isolates from areas where ART resistance is well documented at both temporal and geographical levels. Here we used this strategy to explore the molecular signatures of clinical ART resistance in Cambodia, where this phenotype was first reported^{1,8}.

A candidate molecular marker of ART resistance

The ART-resistant F32-ART5 parasite line was selected by culturing the ART-sensitive F32-Tanzania clone under a dose-escalating, 125-cycle

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regimen of artemisinin for 5 years¹⁸. Whole-genome sequences were obtained for both F32-ART5 and F32-TEM (its sibling clone cultured without artemisinin) at 460× and 500× average nucleotide coverage, respectively. Compared to F32-TEM, no deleted genes were identified in F32-ART5. The exomes of F32-ART5 and F32-TEM were compared after excluding (1) genes from highly variable, multi-gene families (*var*, *rifin* and *stevor*), (2) positions with coverage lower than 25% of the mean coverage of the parasite line, (3) single-nucleotide polymorphisms (SNPs) found to be mixed in F32-ART5, given that acquired ART-resistance mutation(s) could be expected to be fixed in the sample after 5 years of continuous pressure, (4) SNPs shared between F32-ART5 and the ART-sensitive 3D7 parasite strain and (5) synonymous SNPs (Extended Data Fig. 1).

This analysis identified eight mutations in seven genes that were subsequently confirmed by Sanger sequencing of PCR products (Extended Data Table 1). Each gene harbours one mutant codon in F32-ART5 compared to F32-TEM, F32-Tanzania or 3D7 (Extended Data Table 2). Information on the expression of the genes and the biological function of the proteins are listed in Extended Data Table 3. Only one of these genes, cysteine protease falcipain 2a (*PF3D7_1115700*), has previously been associated with *in vitro* responses to ART¹⁹. To determine when each mutation arose in the F32-ART5 lineage, we analysed the whole-genome sequences of parasites at various drug-pressure cycles (Fig. 1). This analysis showed that the *PF3D7_0110400* D56V and *PF3D7_1343700* M476I mutations were acquired first, during the steep increase of ART resistance, and remained stable thereafter. Importantly, the appearance of these two mutations is associated with an increase in the RSA_{0-3h} survival rate, from less than 0.01% to 12.8%. Subsequent PCR analysis of the *PF3D7_1343700* locus detected the M476I mutation after 30 drug-pressure cycles, consistent with the sharp increase in RSA_{0-3h} survival rate observed thereafter. The other SNPs appeared stepwise at later stages of selection: *PF3D7_0213400* (68 cycles); *PF3D7_1115700* (98 cycles); *PF3D7_1302100*, *PF3D7_1459600* and *PF3D7_1464500* (120 cycles) (Extended Data Table 2). These data indicate that the *PF3D7_1343700* M476I mutation increased the resistance of F32-Tanzania to DHA in the RSA_{0-3h}.

To explore whether these mutations are associated with ART resistance in Cambodia, we investigated sequence polymorphism in all seven genes by mining whole-genome or Sanger sequences for 49 culture-adapted parasite isolates collected in 2010–2011 (see Methods). We chose these isolates based on their differential RSA_{0-3h} survival rates (Supplementary Table 1) and their sequences were compared to those of control parasite lines 3D7, 89F5²⁰ and K1992 (see Methods). Three genes (*PF3D7_0110400*, *PF3D7_0213400* and *PF3D7_1302100*) encode a wild-type sequence for all parasite isolates. The other four genes show intra-population diversity, with previously reported or novel SNPs (Supplementary Table 1). *PF3D7_1115700* has 11 SNPs that are not associated with RSA_{0-3h} survival rates ($P = 0.06$, Kruskal–Wallis test). *PF3D7_1459600* has 6 SNPs that are not associated with survival rates ($P = 0.65$). *PF3D7_1464500* has 12 SNPs previously reported in older isolates from southeast Asia, including the ART-susceptible Dd2 line²¹, probably reflecting a geographic signature. These SNPs also show no significant association with survival rates ($P = 0.42$). Therefore, these six genes were not studied further.

In contrast, *PF3D7_1343700* polymorphism shows a significant association with RSA_{0-3h} survival rates (Fig. 2). Indeed, RSA_{0-3h} survival rates differ substantially between parasite isolates with wild-type (median 0.17%, range 0.06–0.51%, $n = 16$) or mutant (18.8%, 3.8–58%, $n = 33$) K13-propeller alleles ($P < 10^{-4}$, Mann–Whitney U test) (Supplementary Table 1). Four mutant alleles are observed, each harbouring a single non-synonymous SNP within a kelch repeat of the C-terminal K13-propeller domain, namely Y493H, R539T, I543T and C580Y located within repeats no. 2, 3, 3 and 4, respectively. Both the K1992 and the ART-susceptible 89F5 lines carry a wild-type K13-propeller. There are no associations between polymorphisms in the K13-propeller and those in the other candidate genes (Supplementary Table 1). Based on these

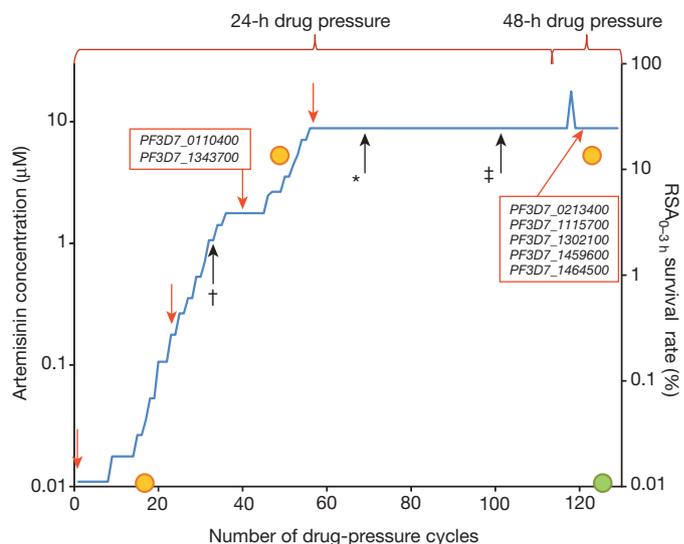


Figure 1 | Temporal acquisition of mutations in F32-ART5. F32-Tanzania parasites exposed to increasing artemisinin concentrations for 120 consecutive cycles¹⁸ were analysed by whole-genome sequencing at five time-points (red arrows). Loci mutated after a given number of drug-pressure cycles are shown (red boxes). The earliest time-points where three mutations were detected by PCR (black arrows) are indicated by † for *PF3D7_1343700*, * for *PF3D7_0213400* and ‡ for *PF3D7_1115700*. Orange and green circles indicate RSA_{0-3h} survival rates for F32-ART5 and F32-TEM parasites, respectively (mean of 3 experiments each).

observations and the acquisition of M476I in kelch repeat no. 2 by F32-ART5, we investigated whether K13-propeller polymorphism is a molecular signature of ART resistance in Cambodia.

Emergence and spread of K13-propeller mutants in Cambodia

Over the last decade, the prevalence of ART resistance has steadily increased in the western provinces of Cambodia, but not elsewhere in

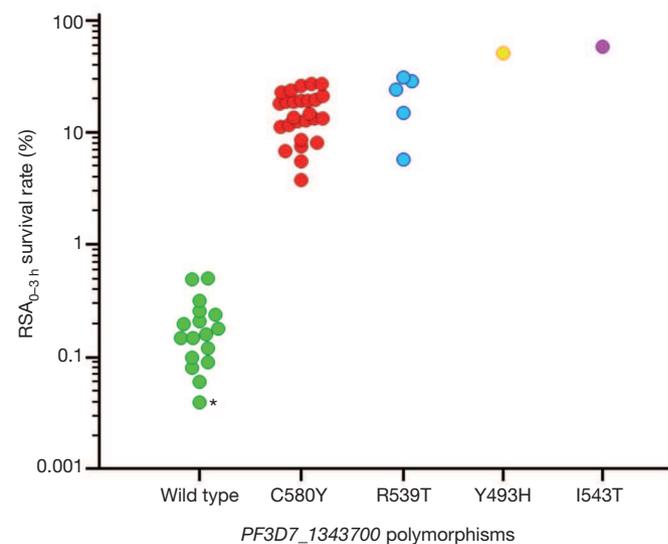


Figure 2 | Survival rates of Cambodian parasite isolates in the RSA_{0-3h} stratified by K13-propeller allele. Genotypes were obtained by mining whole-genome sequence data ($n = 21$) or sequencing PCR products ($n = 28$). Mutant parasites have significantly higher RSA_{0-3h} survival rates than wild-type parasites: wild type ($n = 17$, median 0.16%, IQR 0.09–0.24, range 0.04–0.51); C580Y ($n = 26$, median 14.1%, IQR 11.3–19.6, range 3.8–27.3, $P < 10^{-6}$ for wild type versus C580Y, Mann–Whitney U test); R539T ($n = 5$, median 24.2%, IQR 12.6–29.5, range 5.8–31.3, $P < 10^{-3}$ for wild type versus R539T); Y493H (51.4%); and I543T (58.0%). The RSA_{0-3h} survival rate (0.04%) of control 3D7 parasites is indicated by an asterisk.

the country². To test whether the spatiotemporal distribution of K13-propeller mutations correlates with that of ART resistance, we sequenced the K13-propeller of archived parasite isolates from Cambodian patients with malaria in 2001–2012 (Extended Data Table 4). Data from six provinces were compared ($n = 886$): Pailin, Battambang and Pursat in the west where ART resistance is established^{1,6,8,22}, Kratie in the southeast where ART resistance has increased in recent years², and Preah Vihear in the north and Ratanakiri in the northeast where there was virtually no evidence of ART resistance during this time period². This analysis reveals overall 17 mutant alleles, including three high-frequency (> 5%) alleles (C580Y, R539T and Y493H). The frequency of wild-type sequence decreased significantly over time in all three western provinces, but not in Preah Vihear or Ratanakiri. The frequency of the C580Y allele increased significantly from 2001–2002 to 2011–2012 in Pailin and Battambang, indicating its rapid invasion of the population and near fixation in these areas (Fig. 3).

To further investigate the geographic diversity of K13-propeller polymorphism in Cambodia, we extended our sequence analysis to include data from four additional provinces ($n = 55$, Kampong Som, Kampot, Mondulki and Oddar Meanchey) in 2011–2012 (Extended Data Table 4). Although a large number of mutations are observed (Supplementary Fig. 1 and Extended Data Table 5), the C580Y allele accounts for 85% (189/222) of all mutant alleles observed in 2011–2012 (Extended Data Fig. 2). This mapping outlines the elevated frequency (74%, 222/300) of parasites harbouring a single non-synonymous mutation in the K13-propeller and the geographic disparity of their distribution. Importantly, the frequency distribution of mutant alleles over the various provinces matches that of day 3 positivity in patients treated for malaria with an ACT (Spearman's $r = 0.99$, 95% confidence interval 0.96–0.99, $P < 0.0001$), considered a suggestive sign of clinical ART resistance (Extended Data Fig. 3).

K13-propeller polymorphisms and clinical ART resistance

To confirm that K13-propeller polymorphism is a molecular marker of clinical ART resistance, we first identified 163 patients from Pursat and Ratanakiri in whom we measured parasite clearance half-lives (range 1.58–11.53 h)⁶ in 2009–2010 and for which parasites were previously assigned to a KH subpopulation (KH1, KH2, KH3, KH4 or KHA) on the basis of ancestry analysis of whole-genome sequence data¹⁵. Thirteen patients with mixed genotypes (a wild-type and one or more mutant K13-propeller alleles) were excluded. Of the remaining 150 patients, 72 carried parasites with a wild-type allele and the others carried parasites with only a single non-synonymous SNP in the K13-propeller: C580Y ($n = 51$), R539T ($n = 6$) and Y493H ($n = 21$) (Extended Data Table 6). The parasite clearance half-life in patients with wild-type parasites is significantly shorter (median 3.30 h, interquartile range (IQR) 2.59–3.95) than those with C580Y (7.19 h, 6.47–8.31, $P < 10^{-6}$, Mann-Whitney U test), R539T (6.64 h, 6.00–6.72, $P < 10^{-4}$) or Y493H (6.28 h, 5.37–7.14, $P < 10^{-6}$) parasites (Fig. 4a). Also, the parasite clearance half-life in patients carrying C580Y parasites is significantly longer than those with Y493H parasites ($P = 0.007$, Mann-Whitney U test). These data indicate that C580Y, R539T and Y493H identify slow-clearing parasites in malaria patients treated with ART.

Because KH2, KH3, KH4 and KHA parasites have longer half-lives than KH1 parasites¹⁵, we proposed that allelic variation in the K13-propeller accounts for these differences. Among 150 parasites, 55, 26, 14, 12 and 43 are classified as KH1, KH2, KH3, KH4 and KHA, respectively. Three K13-propeller alleles strongly associate with KH groups: 96% (53/55) of KH1, 96% (25/26) of KH2 and 100% (12/12) of KH4 parasites carry the wild-type, C580Y and Y493H alleles, respectively (Extended Data Table 6). Whereas KH3 parasites ($n = 14$) carry the wild-type, C580Y and R539T alleles, R539T is not observed in KH1, KH2 or KH4

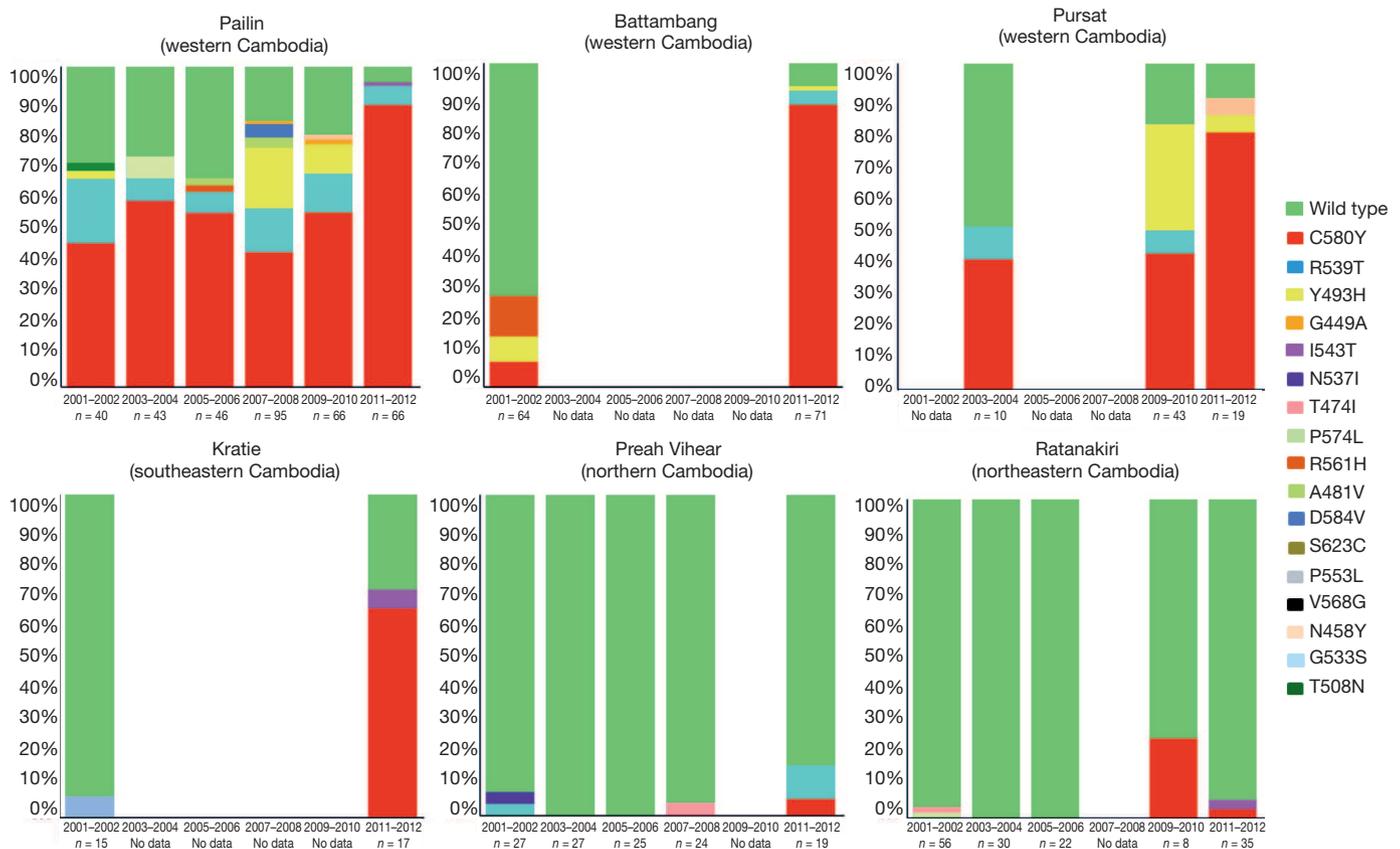


Figure 3 | Frequency of K13-propeller alleles in 886 parasite isolates in six Cambodian provinces in 2001–2012. Genotypes were obtained by sequencing PCR products from archived blood samples. All mutant alleles carry a single non-synonymous SNP (colour-coded, same colour codes as in

Fig. 2 for wild type, C580Y, R539T, Y493H and I543T). Significant reductions (Fisher's exact test) in wild-type allele frequencies were observed in Pailin, Battambang, Pursat and Kratie over time (see Methods).

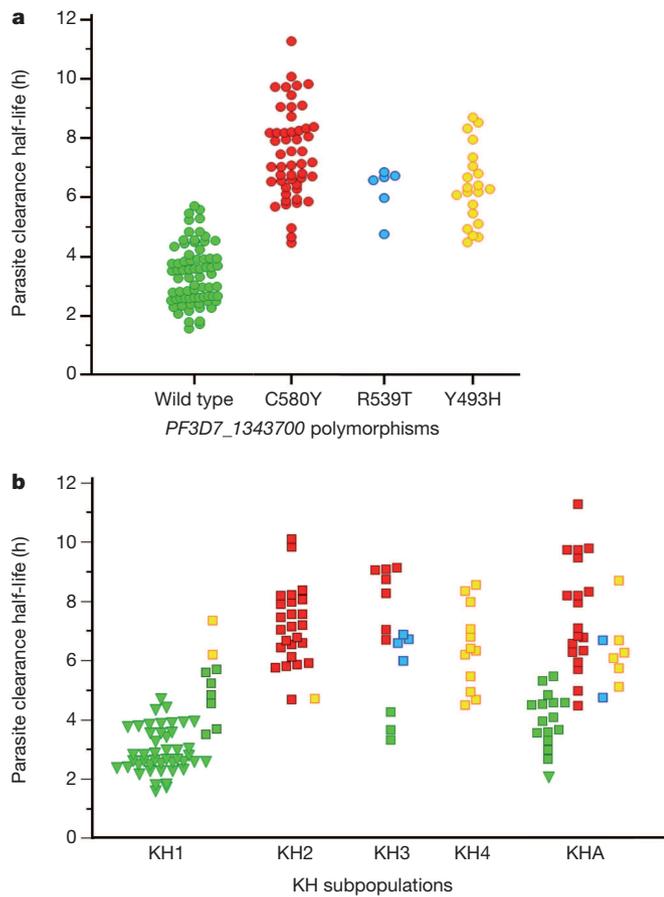


Figure 4 | Parasite clearance half-lives. **a**, Correlation of parasite clearance half-lives and K13-propeller alleles for parasite isolates in Pursat and Ratanakiri in 2009–2010. Wild-type parasites have shorter half-lives (median 3.30 h, IQR 2.59–3.95, $n = 72$) than C580Y (7.19 h, 6.47–8.31, $n = 51$, $P < 10^{-6}$, Mann–Whitney U test), R539T (6.64 h, 6.00–6.72, $n = 6$, $P < 10^{-6}$) or Y493H (6.28 h, 5.37–7.14, $n = 21$, $P < 10^{-6}$) parasites. The half-life of C580Y parasites is significantly longer than that of Y493H parasites ($P = 0.007$). **b**, Correlation of parasite clearance half-lives, KH subpopulations¹⁵ and K13-propeller alleles for the same 150 parasite isolates. Half-lives are shown for Pursat (squares) and Ratanakiri (triangles) parasites, stratified by KH group and K13-propeller allele (colour-coded as in **a**). Median half-lives stratified by K13-propeller allele are KH1: wild type (2.88) and Y493H (6.77); KH2: C580Y (7.13) and Y493H (4.71); KH3: wild type (3.65), C580Y (8.73) and R539T (6.65); KH4: Y493H (6.37); and KHA: wild type (4.01), C580Y (7.09), Y493H (6.18) and R539T (5.73).

parasites. As expected, KHA parasites have a mixed allele composition. Importantly, K13-propeller mutations more accurately identify slow-clearing parasites than KH group (Fig. 4b), demonstrating that the association of K13-propeller polymorphism with clinical ART resistance in Cambodia is partially independent of the genetic background of KH subpopulations. Within the KH1 group ($n = 55$), the parasite clearance half-life in patients with wild-type parasites is significantly shorter ($n = 53$, median 2.88 h, IQR 2.52–3.79) than those with Y493H parasites ($n = 2$, median 6.77 h, $P = 0.02$, Mann–Whitney U test). Within the KH3 subpopulation ($n = 14$), the half-life in patients with wild-type parasites is shorter ($n = 3$, median 3.65 h) than those with C580Y ($n = 7$, median 8.73 h, IQR 7.35–9.06, $P = 0.02$) or R539T ($n = 4$, 6.65 h, 6.29–6.80, $P = 0.03$) parasites.

Discussion

The F32-ART5 lineage acquired a K13-propeller mutation as it developed ART resistance, as indicated by its ability to survive a pharmacologically relevant exposure to DHA in the RSA_{0–3h}. Genes putatively associated with ART resistance (*Pfcr*^{23,24}, *Pftct*^{25,26}, *Pfmdr1*^{8,27,28}, *Pfmrp1*^{27–29} and

ABC transporters³⁰) or encoding putative targets of ART (*PfATPase6*^{31,32} and *Pfubctb*—the orthologue of *Plasmodium chabaudi* ubp1^{33,34}) were not mutated during the 5-year selection of F32-ART5, and *Pfmdr1* amplification was not observed^{35–40}. In addition, all candidate ART-resistance genes recently identified using population genetics approaches^{14,40,41} remained unaltered in F32-ART5, except for *PF3D7_1343700* and *PF3D7_1459600* located in the linkage-disequilibrium windows identified in ref. 16. These findings led us to identify another 17 single K13-propeller mutations in naturally circulating parasites in Cambodia. Several of these mutations associate strongly with the spatiotemporal distribution of ART resistance in Cambodia, increased parasite survival rates in response to DHA *in vitro*, and long parasite clearance half-lives in response to ART treatment *in vivo*. None of the six other genes mutated in F32-ART5 associate with RSA_{0–3h} survival rates in parasite isolates from Cambodia.

K13-propeller polymorphism fulfils the definition of a molecular marker of ART resistance for several reasons: (1) there has been a progressive loss of wild-type parasites in western Cambodia during the decade of emerging ART resistance in this region; (2) mutant parasites cluster in Cambodian provinces where ART resistance is well established and are less prevalent where ART resistance is uncommon; (3) *PF3D7_1343700* is located 5.9 kilobases upstream of the 35-kb locus identified in ref. 14 as being under recent positive selection, and within the region of top-ranked signatures of selection outlined in ref. 16; (4) multiple mutations, all non-synonymous, are present in the K13-propeller, reflecting positive selection rather than a hitchhiking effect or genetic drift; (5) mutations occur in a domain that is highly conserved in *P. falciparum*, with only one non-synonymous SNP being documented in a single parasite isolate from Africa⁴²; (6) all polymorphisms we observe in Cambodia are novel and all but one (V568G) occur at positions strictly conserved between *Plasmodium* species (Supplementary Fig. 1 and Supplementary Fig. 2), suggesting strong structural and functional constraints on the protein; (7) the three most-prevalent K13-propeller mutations correlate strongly with RSA_{0–3h} survival rates *in vitro* and parasite clearance half-lives *in vivo* at the level of individual parasite isolates and malaria patients, respectively; and (8) the frequency of mutant alleles correlates strongly with the prevalence of day 3 positivity after ACT treatment at the level of human populations in Cambodia.

On the basis of homology with other kelch propeller domains, we anticipate that the observed K13-propeller mutations destabilize the domain scaffold and alter its function. The carboxy-terminal portion of *PF3D7_1343700* encodes six kelch motifs, which are found in a large number of proteins with diverse cellular functions^{43,44}. Given that the toxicity of ART derivatives depends principally on their pro-oxidant activity, the reported role of some kelch-containing proteins in regulating cytoprotective and protein degradation responses to external stress is particularly intriguing. The K13-propeller shows homology with human KLHL12 and KLHL2, involved in ubiquitin-based protein degradation, and KEAP1, involved in cell adaptation to oxidative stress (Extended Data Fig. 4). More work is needed to delineate the normal function of K13 and the effect of various mutations. Allele exchange studies in mutant and wild-type parasites may help to define the contribution of K13-propeller polymorphisms on different genetic backgrounds to the RSA_{0–3h} survival rate. Indeed, it is particularly worrying that as few as two mutations, that is, the K13-propeller M476I and *PF3D7_0110400* D56V, were sufficient to confer ART resistance to F32-Tanzania, which has a typical African genetic background. Cambodian parasites with mutant K13-propellers display a wide range of RSA_{0–3h} survival rates (3.8–58%) and parasite clearance half-lives (4.5–11.5 h). Further studies are therefore required to identify additional genetic determinants of ART resistance, which may reside in the strongly selected regions recently identified^{14,16}. In this context, analysing the RSA_{0–3h} survival rates as a quantitative trait among parasites harbouring the same K13-propeller mutation could help to identify additional genetic loci involved in ART resistance.

In summary, K13-propeller polymorphism seems to be a useful molecular marker for tracking the emergence and spread of ART-resistant *P. falciparum*.

METHODS SUMMARY

The ART-resistant F32-ART5 parasite line was selected by culturing the ART-sensitive F32-Tanzania clone under a dose-escalating regimen of artemisinin for 5 years. The F32-TEM line was obtained by culturing F32-Tanzania in parallel without artemisinin exposure. Reference DNA was extracted from *P. falciparum* lines 3D7, 89F5 Palo Alto Uganda and K1992. The ring-stage survival assay (RSA_{0-3h}) was performed as described previously¹³. Whole-genome sequencing was performed on F32-Tanzania, F32-TEM, F32-ART5 (4 time points), three reference strains (3D7, 89F5 and K1992) and 21 Cambodian parasite isolates, using an Illumina paired-reads sequencing technology. A set of 1091 clinical *P. falciparum* isolates was collected from patients participating in ACT efficacy studies in 2001–2012. The K13-propeller was amplified using nested PCR. Double-strand sequencing of PCR products was performed by Macrogen. Sequences were analysed with MEGA 5 software version 5.10 to identify specific SNP combinations. Data were analysed with Microsoft Excel and MedCalc version 12. Differences were considered statistically significant when *P* values were less than 0.05. Ethical clearances for parasite isolate collections were obtained from the Cambodian National Ethics Committee for Health Research, the Institutional Review Board of the Naval Medical Research Center, the Technical Review Group of the WHO Regional Office for the Western Pacific, and the Institutional Review Board of the National Institute of Allergy and Infectious Diseases.

Online Content Any additional Methods, Extended Data display items and Source Data are available in the online version of the paper; references unique to these sections appear only in the online paper.

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and D.M. conceived of the study, supervised the project, processed the data and wrote the manuscript with contributions from B.W., C.A., A.B. and J.-C.B.

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METHODS

Artemisinin- and mock-pressured *P. falciparum* F32 lineages. Mycoplasma-free F32-Tanzania parasites were maintained in human type O red blood cells (RBCs) (Etablissement Français du Sang) diluted to 2.5% haematocrit in RPMI-1640 medium (Invitrogen, San Diego, CA) supplemented with 5% human serum. Parasite cultures were maintained at 37 °C in an atmosphere of 5% CO₂. Parasitaemia was checked daily and maintained below 10%. For the selection of ART-resistant parasites, asynchronous cultures were adjusted to 5–7% parasitaemia and grown in the presence of escalating doses of artemisinin (from 10 nM to 9 μM) for 24 h for the first 3 years of drug pressure¹⁸. In the subsequent 2 years, each drug-pressure cycle was done for 48 h with doses ranging from 9 μM to 18 μM. After drug exposure, the medium was discarded and replaced by human-serum-supplemented (20%) drug-free medium. Parasitaemia was monitored daily until it reached 5%. At that time, drug pressure was reapplied. The parasite line obtained after an effective 5 years of discontinuous ART pressure was named F32-ART5. In parallel, the parental F32-Tanzania line was kept as a control in continuous culture for the same time under the same conditions (that is, RBCs, serum and media) but without artemisinin exposure. The resulting control line was called F32-TEM.

Laboratory-adapted *P. falciparum* lines. Reference DNA was extracted from the laboratory-adapted *P. falciparum* lines 3D7 (MR4, Manassas, VA), 89F5 Palo Alto Uganda (a clone from the Palo Alto line, originating from Uganda in 1978, which displayed high susceptibility to artemether treatment in the *Saimiri sciureus* experimental host (O. Mercereau-Puijalon, H. Contamin and J.-C. Barale, unpublished data)) and K1992, an isolate collected in Pailin in 1992 before the mass deployment of ART in that area (provided by the French National Reference Center of Malaria). Parasite DNA was extracted from frozen blood aliquots (200 μl) using the Mini blood kit (Qiagen) according to the manufacturer's instructions.

Culture-adapted *P. falciparum* isolates from Cambodia. Fifty clinical *P. falciparum* isolates from Cambodia (collected in 2010 and 2011) were adapted to *in vitro* culture as described in ref. 45. Their geographic origin is indicated in Supplementary Table 1. Parasite clearance rates were not determined for these patient isolates, as they were collected during field trials that did not include such measurements. Parasite DNA was extracted from frozen blood aliquots (200 μl) using the Mini blood kit (Qiagen).

Ring-stage survival assay. The ring-stage survival assay (RSA_{0–3h}) was carried out as described in ref. 13 using highly synchronous parasite cultures. In brief, 0–3 h post-invasion ring-stage parasites were exposed to 700 nM DHA (dihydroartemisinin, obtained from WWARN (<http://www.wwarn.org/research/tools/qaqc>)) or its solvent DMSO for 6 h, washed and then cultivated for the next 66 h without drug. Survival rates were assessed microscopically by counting in Giemsa-stained thin smears the proportion of viable parasites that developed into second-generation rings or trophozoites with normal morphology.

Ethical clearance. Ethical clearances for the collection of parasite isolates from patients were obtained from the Cambodian National Ethics Committee for Health Research, the Institutional Review Board of the Naval Medical Research Center, the Technical Review Group of the WHO Regional Office for the Western Pacific, and the Institutional Review Board of the National Institute of Allergy and Infectious Diseases. Work was conducted in compliance with all relevant ethical standards and regulations governing research involving human subjects. Written informed consent was obtained from all adult participants or the parents or guardians of children.

Temporal and geographical sample collection. A set of 941 clinical *P. falciparum* isolates was collected from patients participating in therapeutic efficacy studies of ACTs, conducted as part of the routine antimalarial drug efficacy monitoring of Cambodia's National Malaria Control Program from 2001 to 2012, and from studies conducted by NAMRU-2 (Extended Data Table 4). Venous blood samples (5 ml) collected in EDTA or ACD were transported to Institut Pasteur du Cambodge in Phnom Penh within 48 h of collection at 4 °C and then kept at –20 °C until DNA extraction. Parasite DNA was extracted from frozen blood aliquots (200 μl) using the Mini blood kit (Qiagen).

Measurement of parasite clearance half-life. Patients with uncomplicated or severe *P. falciparum* malaria and initial parasite density $\geq 10,000 \mu\text{l}^{-1}$ were enrolled in Pursat and Ratanakiri provinces in 2009 and 2010 as described^{6,13}. Patients were treated with an ART and their parasite density measured every 6 h from thick blood films until parasitaemia was undetectable. The parasite clearance half-life in 163 patients was derived from these parasite counts using WWARN's on-line Parasite Clearance Estimator (<http://www.wwarn.org/toolkit/data-management/parasite-clearance-estimator>). The study is registered at ClinicalTrials.gov (number NCT00341003).

Whole-genome sequencing of parasite DNA. Whole-genome sequencing was performed on F32-Tanzania, F32-TEM, the F32-ART5 lineage (4 time-points), three reference strains (3D7, 89F5 and K1992) and 21 parasite isolates from Cambodia, using an Illumina paired-reads sequencing technology. Illumina library preparation

and sequencing followed standard protocols developed by the supplier. Briefly, genomic DNA was sheared by nebulization, and sheared fragments were end-repaired and phosphorylated. Blunt-end fragments were A-tailed, and sequencing adapters were ligated to the fragments. Inserts were sized using Agencourt AMPure XP Beads (± 500 bp; Beckman Coulter Genomics) and enriched using 10 cycles of PCR before library quantification and validation. Hybridization of the library to the flow cell and bridge amplification was performed to generate clusters, and paired-end reads of 100 cycles were collected on a HiSeq 2000 instrument (Illumina). After sequencing was complete, image analysis, base calling and error estimation were performed using Illumina Analysis Pipeline version 1.7.

Raw sequence files were filtered using Fqquality tool, a read-quality filtering software developed by N. Joly, which enables the trimming of the first and last low-quality bases in reads. The trimmed reads from controlled Fastq files were mapped on a reference genome (*P. falciparum* 3D7) with the Burrows-Wheeler Alignment (BWA), generating a BAM file (a binary file of tab-delimited format SAM). Next, we used Samtools to prepare a pileup file, which was formatted using in-house software to implement the data into the Wholegenome Data Manager (WDM) database (Beghain *et al.*, in preparation). WDM software is designed to compare and/or align partial or whole *P. falciparum* genomes.

Sequencing genes containing non-synonymous SNPs in F32-ART5. PCR amplification of selected genes was performed using the primers listed in Extended Data Table 1. Two μl of DNA was amplified with 1 μM of each primer, 0.2 mM dNTP (Solis Biodyne), 3 mM MgCl₂ and 2 U *Taq* DNA polymerase (Solis Biodyne), using the following cycling program: 5 min at 94 °C, then 40 cycles of 30 s at 94 °C, 90 s at 60 °C, 90 s at 72 °C and final extension 10 min at 72 °C. PCR products were detected by 2% agarose gel electrophoresis and ethidium bromide staining. Double-strand sequencing of PCR products was performed by Beckman Coulter Genomics. Sequences were analysed with MEGA 5 software version 5.10 in order to identify specific SNP combinations.

Sequencing the K13-propeller domain. The K13-propeller domain was amplified using the following primers: for the primary PCR (K13-1 5'-cggagtaccacaaactggga-3' and K13-4 5'-gggaatctggtgtaacagc-3') and the nested PCR (K13-2 5'-gccaaagctgcattcattg-3' and K13-3 5'-gcctgtgaaagaagcaga-3'). One μl of DNA was amplified with 1 μM of each primer, 0.2 mM dNTP (Solis Biodyne), 3 mM MgCl₂ and 2 U *Taq* DNA polymerase (Solis Biodyne), using the following cycling program: 5 min at 94 °C, then 40 cycles of 30 s at 94 °C, 90 s at 60 °C, 90 s at 72 °C and final extension 10 min at 72 °C. For the nested PCR, 2 μl of primary PCR products were amplified under the same conditions, except for the MgCl₂ concentration (2.5 mM). PCR products were detected using 2% agarose gel electrophoresis and ethidium bromide staining. Double-strand sequencing of PCR products was performed by Macrogen. Sequences were analysed with MEGA 5 software version 5.10 to identify specific SNP combinations.

Deep-sequencing of clinical parasite isolates and population structure analysis. DNA extraction, Illumina sequencing and SNP genotyping of clinical parasite isolates obtained from malaria patients in Pursat and Ratanakiri provinces, Cambodia, have been previously described¹⁵. Population structure analysis of these parasites identified four subpopulations: KH1, KH2, KH3 and KH4. Parasites with <80% ancestry from any of these four groups were deemed admixed (KHA).

Temporal acquisition of mutations in the F32-ART5 lineage. The F32-ART5 lineage was explored by whole-genome sequencing using samples collected at time 0 (original F32-Tanzania clonal line), day 196 (0.2-μM pressure cycle no. 23), day 385 (1.8-μM pressure cycle no. 39), day 618 (9-μM pressure cycle no. 56) and day 2,250 (9-μM pressure cycle no. 120). The F32-TEM sample was collected on day 2,250. Additional samples collected at the time of the 30th, 33rd, 34th, 36th, 68th and 98th pressure cycles were studied by PCR. DNA from parasite cultures was extracted using the High Pure PCR Template Preparation Kit (Roche Diagnostics) according to the manufacturer's instructions.

The F32-ART5 samples tested in the ring-stage survival assay (RSA_{0–3h}) were collected at the time of the 17th, 48th and 122nd pressure cycles (0.04, 2.7 and 9 μM ART), respectively. The F32-TEM sample was collected at the last mock pressure cycle. The RSA_{0–3h} survival rates were determined in triplicate experiments with different batches of red blood cells, and evaluated as above using Giemsa-stained thin smears read by two independent microscopists (B.W. and F.B.-V.). Survival rates were compared using Mann-Whitney *U* test. The RSA_{0–3h} survival rates of the F32-ART5 samples were as follows: at drug-pressure cycles: no. 17 ($n = 3$, median 0%, IQR 0–0.07), no. 48 ($n = 3$, median 11.7%, IQR 10.3–14.6; $P = 0.04$ for no. 17 versus no. 48, Mann-Whitney *U* test) and no. 122 ($n = 3$, median 12.8%, IQR 10.6–14.5, $P = 0.04$ and $P = 0.82$ for no. 17 versus no. 122 and no. 48 versus no. 122). The RSA_{0–3h} survival rate of the F32-TEM line was also determined in triplicate experiments ($n = 3$, median 0%, IQR 0–0.05, $P = 0.81$ for TEM versus no. 17, Mann-Whitney *U* test).

Prevalence of K13-propeller mutations in 886 clinical parasite isolates collected in six Cambodian provinces in 2001–2012. The K13-propeller was genotyped by

sequencing PCR products amplified from 886 archived blood samples. The number of samples analysed from each province each year is indicated in Fig. 3. Fisher's exact test was used to compare the frequency of isolates harbouring a wild-type K13-propeller sequence in each province over time. A significant decrease of the frequency of the wild-type K13-propeller allele was observed in the western provinces during the decade. In Pailin, it decreased from 30.0% in 2001–2002 (12/40) to 4.8% in 2011–2012 (4/84), $P = 0.0002$, in Battambang from 71.9% in 2001–2002 (46/64) to 7.0% in 2011–2012 (5/71), $P < 10^{-6}$, in Pursat from 50.0% in 2003–2004 (5/10) to 10.5% in 2011–2012 (2/19), $P = 0.03$; and in Kratie from 93.3% in 2001–2002 (14/15) to 29.4% in 2011–2012 (5/17), $P = 0.0003$. Significant decreases in wild-type allele frequency were not observed in Preah Vihear (from 92.6% in 2001–2002 (25/27) to 84.2% in 2011–2012 (16/19), $P = 0.63$); or Ratanakiri (from 96.4% in 2001–2002 (54/56) to 94.3% in 2011–2012 (33/35), $P = 0.63$). The frequency of C580Y increased in Pailin from 45.0% (18/40) in 2001–2002 to 88.1% (74/84) in 2011–2012 ($P < 10^{-6}$), and in Battambang from 7.8% (5/64) in 2001–2002 to 87.3% (62/71) in 2011–2012 ($P < 10^{-6}$) indicating its rapid invasion of the population and near fixation in these provinces.

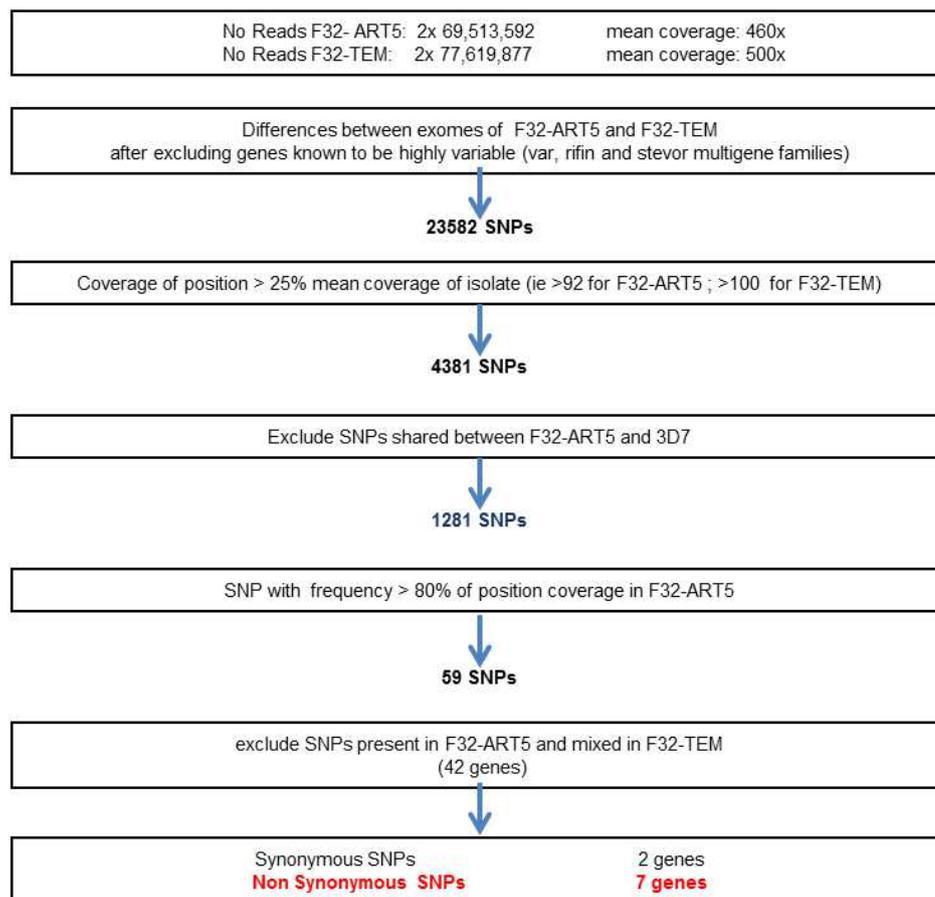
Three-dimensional structure modelling of the K13-propeller. The 3D-structural model of the kelch propeller domain of *PF3D7_1343700* ('K13-propeller') was obtained by homology modelling satisfying spatial restraints using Modeller v9.11 (<http://modbase.compbio.ucsf.edu>). The 295 amino acids composing the K13-propeller are 22%, 25% and 25% identical to the kelch propeller domain of the human KEAP1 (Protein Data Bank (PDB; <http://www.rcsb.org/>) 2FLU), KLHL12 (PDB 2VPJ) and KLHL2 (PDB 2XN4) proteins, respectively, that were used as templates to model the 3D-structure of the K13-propeller. The reliability of the obtained model was assessed using two classical criteria. First, the significance of the sequence alignment between the K13-propeller domain and one template was confirmed by an E -value = 0, as calculated by Modeller using the Built-Profile routine. Second, the model achieved a GA341 model score = 1 (a score ≥ 0.7 corresponds to highly reliable models). Localization of the mutants in the K13-propeller 3D-model was prepared using the PyMOL Molecular Graphics System, version 1.5.0.4 (Schrödinger; <http://www.pymol.org>).

Statistical analysis. Data were analysed with Microsoft Excel and MedCalc version 12. Quantitative data were expressed as median, interquartile range (IQR). The Mann–Whitney U test (independent samples, two-sided) was used to compare two groups, and the Kruskal–Wallis test (H -test, two-sided) was used to compare more than two groups. The Spearman's rho rank correlation coefficient (and the 95% confidence interval for the correlation coefficient) was used to measure the strength of relationship between the prevalence of wild-type K13-propeller allele and the frequency of day 3 positivity (defined as persistence of microscopically detectable parasites on the third day of artemisinin-based combination therapy)². Fisher's exact test was used to compare frequency data and the Clopper–Pearson exact method based on the beta distribution was used to determine

95% confidence intervals for proportions. Differences were considered statistically significant when P values were less than 0.05.

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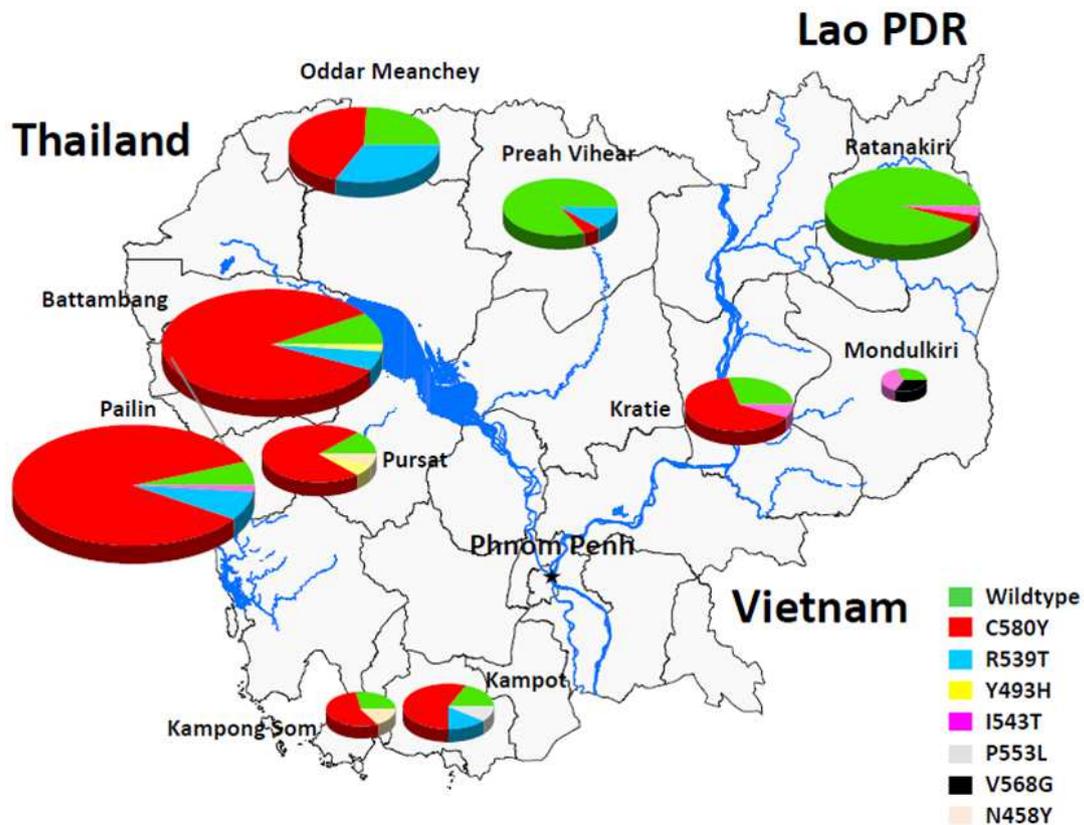
a



b

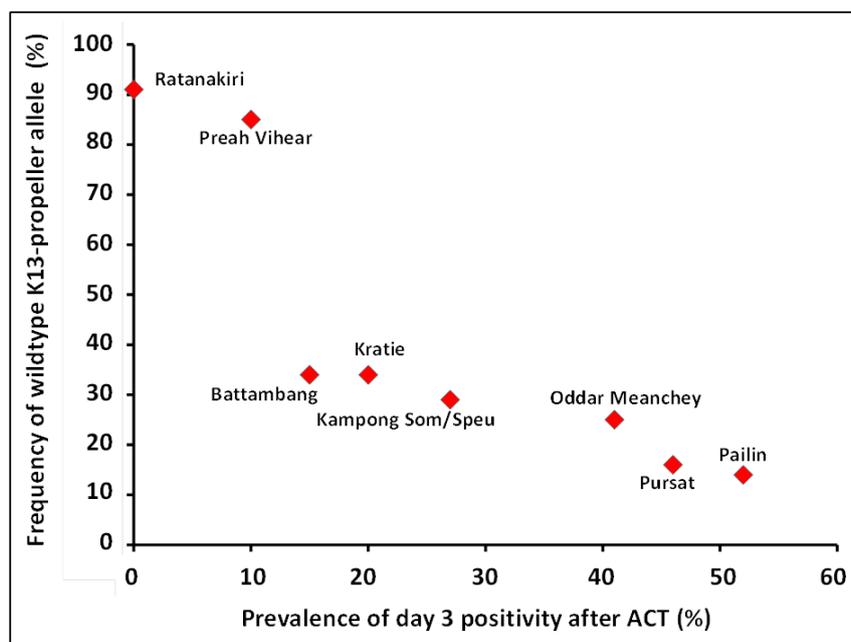
Chromosome	Position	Gene ID	F32-ART5			F32-TEM		
			coverage	No reads with mutant SNP	% reads with mutant SNP	coverage	No reads with w.t. SNP	% reads with w.t. SNP
1	394452	PF3D7_0110400;DNA-directed RNA polymerase 2, putative	224	222	99.11	335	334	99.70
2	542625	PF3D7_0213400;protein kinase 7 (PK7)	242	242	100.00	403	403	100.00
11	593379	PF3D7_1115700;cysteine proteinase falcipain 2a	234	231	98.72	290	289	99.66
13	121689	PF3D7_1302100;gamete antigen 27/25 (Pfg27)	261	259	99.23	343	342	99.71
13	1725570	PF3D7_1343700;kelch protein, putative	1004	1004	100.00	1161	1160	99.91
14	2442240	PF3D7_1459600;conserved Plasmodium protein, unknown	165	142	86.06	225	225	100.00
14	2612177	PF3D7_1464500;conserved Plasmodium membrane protein, unknown	401	399	99.50	428	428	100.00

Extended Data Figure 1 | SNP-calling algorithm and sequence and coverage of SNPs. a, SNP-calling algorithm of the whole-genome sequence comparison of F32-ART5 and F32-TEM. b, Sequence and coverage of SNPs in seven candidate genes differing in F32-TEM and F32 ART5.



Extended Data Figure 2 | Geographic distribution of K13-propeller alleles in Cambodia in 2011–2012. Pie charts show K13-propeller allele frequencies among 300 parasite isolates in ten Cambodian provinces. Pie sizes are proportional to the number of isolates and the different alleles are colour-coded as indicated. The frequencies (95% confidence interval) of mutant K13-

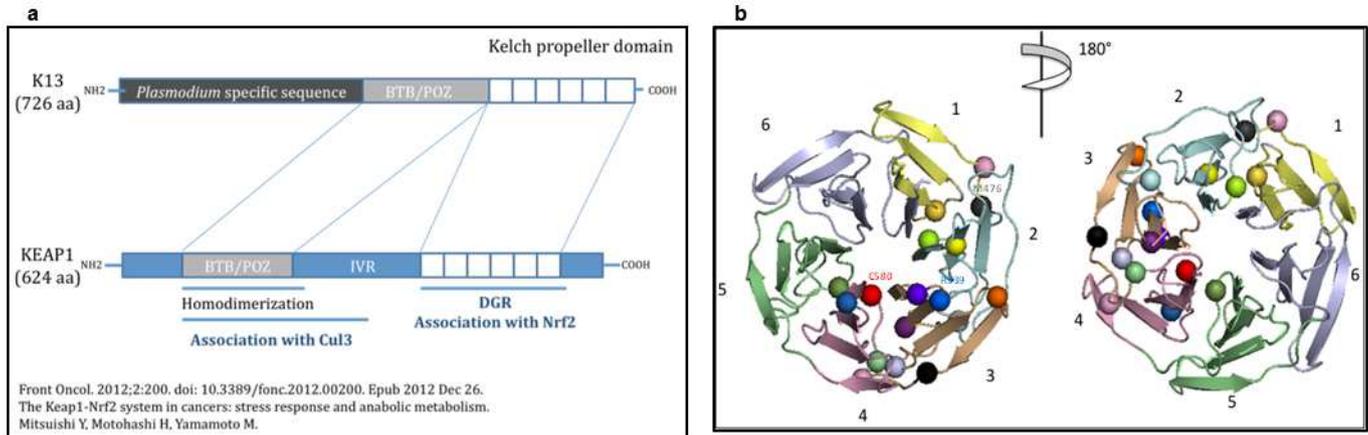
propeller alleles are: Pailin (95%, 88–99, $n = 84$), Battambang (93%, 87–99, $n = 71$), Pursat (89%, 67–99, $n = 19$), Kampot (83%, 52–98, $n = 12$), Kampong Som (71%, 29–96, $n = 7$), Oddar Meanchey (76%, 58–89, $n = 33$), Preah Vihear (16%, 3–40, $n = 19$), Kratie (71%, 44–90, $n = 17$), Monduliri (67%, 9–99, $n = 3$) and Ratanakiri (6%, 1–19, $n = 35$).



Extended Data Figure 3 | Correlation between the frequency of wild-type K13-propeller alleles and the prevalence of day 3 positivity after ACT treatment in eight Cambodian provinces.

The frequency of day 3 positivity is plotted against the frequency of wild-type K13-propeller alleles. Data are derived from patients treated with an ACT for *P. falciparum* malaria in 2010–2012 in eight Cambodian provinces (Extended Data Figure 2): Pailin ($n = 86$, 2011 WHO therapeutic efficacy study, artesunate-mefloquine); Pursat ($n = 32$, 2012 WHO therapeutic efficacy study, dihydroartemisinin-piperaquine); Oddar Meanchey ($n = 32$, 2010 NAMRU-2 therapeutic efficacy study,

artesunate-mefloquine); Kampong Som/Speu ($n = 7$, 2012 WHO therapeutic efficacy study, dihydroartemisinin-piperaquine); Battambang ($n = 18$, 2012 WHO therapeutic efficacy study, dihydroartemisinin-piperaquine); Kratie ($n = 15$, 2011 WHO therapeutic efficacy study, dihydroartemisinin-piperaquine); Preah Vihear ($n = 19$, 2011 WHO therapeutic efficacy study, dihydroartemisinin-piperaquine); Ratanakiri ($n = 32$, 2010 WHO therapeutic efficacy study, dihydroartemisinin-piperaquine). Spearman's coefficient of rank correlation (8 sites): $r = -0.99$, 95% confidence interval -0.99 to -0.96 , $P < 0.0001$.



Extended Data Figure 4 | Schematic representation of homology between *P. falciparum* K13 and human KEAP1 proteins and structural 3D model of the K13-propeller domain. **a**, Schematic representation of the predicted *PF3D7_1343700* protein and homology to human KEAP1. Similar to KEAP1, *PF3D7_1343700* contains a BTB/POZ domain and a C-terminal 6-blade propeller, which assembles kelch motifs consisting of four anti-parallel beta sheets. **b**, Structural 3D model of the K13-propeller domain showing the six kelch blades numbered 1 to 6 from N to C terminus and colour-coded as in Supplementary Fig. 1. The level of amino-acid identity between the K13-propeller and kelch domains of proteins with solved 3D structures, including human KEAP1^{46,47}, enabled us to model the 3D structure of the K13-propeller and to map the mutations selected under ART pressure (Extended data Table 5). The accuracy of the K13-propeller 3D model was confirmed by Modeller-specific model/fold criteria of reliability (see Methods). We predict that the K13-propeller folds into a 6-bladed β -propeller structure⁴⁸ closed by the interaction between a C-terminal beta-sheet and the N-terminal blade^{46,48}. The first domain has three β -sheets, the fourth one being contributed by an extra C-terminal β -sheet called $\beta'1$ in Supplementary Fig. 1. The human KEAP1 kelch propeller scaffold is destabilized by a variety of mutations affecting intra- or inter-blade interactions in human lung cancer⁴⁶ and hypertension⁴⁷. The positions of the various mutations are indicated by a

sphere, colour-coded as in Figs 2–4. The M476 residue mutated in F32-ART5 is indicated in dark grey. Like the mutations observed in human KEAP1^{46,47}, many K13-propeller mutations are predicted to alter the structure of the propeller or modify surface charges, and as a consequence alter the biological function of the protein. Importantly, the two major mutations C580Y (red) and R539T (blue) observed in Cambodia are both non-conservative and located in organized secondary structures: a β -sheet of blade 4 where it is predicted to alter the integrity of this scaffold and at the surface of blade 3, respectively. The kelch propeller domain of KEAP1 is involved in protein–protein interactions like most kelch containing modules⁴³. KEAP1 is a negative regulator of the inducible Nrf2-dependent cytoprotective response, sequestering Nrf2 in the cytoplasm under steady state. Upon oxidative stress, the Nrf2/KEAP1 complex is disrupted, and Nrf2 translocates to the nucleus, where it induces transcription of cytoprotective ARE-dependent genes^{49,50}. We speculate that similar functions may be performed by *PF3D7_1343700* in *P. falciparum*, such that mutations of the K13-propeller impair its interactions with an unknown protein partner, resulting in a deregulated anti-oxidant/cytoprotective response. The *P. falciparum* anti-oxidant response is maximal during the late trophozoite stage, when haemoglobin digestion and metabolism are highest⁵¹. Its regulation is still poorly understood and no Nrf2 orthologue could be identified in the *Plasmodium* genome.

Extended Data Table 1 | Sequence of the primers used to amplify the genes containing nonsynonymous single-nucleotide polymorphisms in F32-ART5

Targeted gene	Primer forward sequence	Primer reverse sequence
PF3D7_0110400	5'-ttgagcttctttttoccaataatggc-3'	5'-tgataatgtttgtaggagctgtgag-3'
PF3D7_0213400	5'-gtgaaaaggataataaattctatgcc-3'	5'-tatctaccatataattctgattctcc-3'
PF3D7_1115700	5'-agcaagaacgttttgggtgtaa-3'	5'-gaattotttaatggtttgaagat-3'
PF3D7_1302100	5'-taatatgtaaagtgattatgtatatcgc-3'	5'-atgctagagaagttaagagaagaagcg-3'
PF3D7_1343700	5'-agaagagccatcatatccccc-3'	5'-agtggaagacatcatgtaaccag-3'
PF3D7_1459600	5'-atatgagtaaaatgacaggttttgg-3'	5'-tgcttgtgtgattcatgggg-3'
PF3D7_1464500	5'-aaatagttggcgtagctcag-3'	5'-tatcacaattaagtgtatcacaacg-3'

Extended Data Table 2 | Description of the eight nonsynonymous, single-nucleotide polymorphisms acquired in the F32-ART5 compared to the F32-TEM lineage during an effective 5-year discontinuous exposure to increasing concentrations of artemisinin

Gene ID (Plasmodb 9.1)	Annotation	Chromosome/ position mutated	Nucleotide position in coding sequence	F32- TEM codon ^a	Codon F32-ART5 lineage					Mutant codon
					Day 0	Drug pressure cycle #				
						23	39	56	120	
					-	0.2 μ M ART [*]	1.8 μ M ART [*]	9 μ M ART [*]	9 μ M ART [*]	
PF3D7_0110400	DNA-directed RNA polymerase 2 complex subunit RPB9, putative	01/39452	173	gAt	gAt	gAt	gTg	gTg	gTg	D56V
PF3D7_1343700 ^a	kelch protein, putative, called here 'K13'	13/1725570	1428	atG	atG	atG	atA	atA	atA	M476I
PF3D7_0213400	protein kinase 7 (PK7)	02/542625 02/542627	310 312	GaA	GaA	GaA	GaA	GaA	TaG	E104stop
PF3D7_1115700	cysteine proteinase falcipain 2a	11/593378	206	tCa	tCa	tCa	tCa	tCa	tGa	S69stop
PF3D7_1302100	gamete antigen 27/25 (Pfg27)	13/121689	601	Cca	Cca	Cca	Cca	Cca	Aca	P201T
PF3D7_1459600 ^a	conserved <i>Plasmodium</i> protein, unknown function	14/2442240	896	aGt	aGt	aGt	aGt	aGt	aCt	S299T
PF3D7_1464500	conserved <i>Plasmodium</i> membrane protein, unknown function	14/2612177	4886	aAt	aAt	aAt	aAt	aAt	aGt	N1629S

3D7-type sequence; the same codon sequence is also observed in the parental F32-Tanzania line.

* Artemisinin (ART) dose used for selection during the corresponding drug-pressure cycle.

^aGenes found in the chromosomal location of top-ranked signatures of selection in ref. 16.

Extended Data Table 3 | Reported characteristics of the genes mutated in F32-ART5 parasites

PF3D7_0110400 (PFA0505c), is a two-exon gene, codes for the RNA Polymerase II subunit 9 (RPB9), a small integral Pol II subunit, which is highly conserved among eukaryotes. The yeast RPB9 ortholog has been shown to have a role in assuring the fidelity of transcription *in vivo*. Deletion of the gene results in error-prone transcription⁵². The protein has a predicted zinc ribbon domain similar to the zinc ribbon domain of TFIIS (RNA Polymerase II elongation factor) that contains the essential catalytic Asp-Glu dipeptide⁵³. Very little is known on the protein in *Plasmodia*, although the gene is expressed and the protein is present in blood stages (www.plasmodb.org). It is difficult to make any prediction on the possible phenotypic consequences of the D56V mutation, which is located in a *Plasmodium*-specific, well-conserved domain.

PF3D7_1343700 (PF13_0238), is a one-exon gene (called here K13) that codes for a putative kelch protein. K13 has a predicted 3-domain structure, with an approx. 225 residue long, *Plasmodium*-specific and well conserved N-terminal domain, followed by a BTB/ POZ domain and a 6-blade C-terminal propeller domain formed of canonical kelch motifs^{43,48}. Little is known about the protein in malaria parasites. Proteomics data indicate that it is produced by asexual (trophozoites, schizonts, merozoites and rings) and sexual blood stages (gametocytes) of *P. falciparum*, and that it possesses phosphorylated residues in the N-terminal *Plasmodium*-specific domain (www.plasmodb.org). The M476I mutation is located between the first and second blade of the propeller domain.

PF3D7_0213400 (PFB0605w), is a four-exon gene that codes for protein kinase 7 (PK7) expressed during the asexual blood stage development, in gametocytes and ookinetes. The E104 stop mutation (two SNPs affecting the same codon) observed in F32-ART5 interrupts the gene resulting in a truncated putative translation product lacking more than 2/3 of its sequence. Studies with genetically inactivated parasites have shown that PK7-KO *P. falciparum* parasites have an asexual growth defect due to a reduced number of merozoites per schizont⁵⁴. Furthermore, PK7 is important for mosquito transmission, with a collapsed number of ookinetes in *P. falciparum*⁵⁴ and in *P. berghei*, where no sporoblasts and consequently no sporozoites are formed⁵⁵. This transmission defective phenotype is unlikely to survive in the field.

PF3D7_1115700 (PF11_0165), is a one-exon gene that codes for falcipain 2a, a cysteine proteinase produced by maturing blood stages (trophozoites and schizonts) and involved in hemoglobin degradation⁵⁶. The S69stop mutation located in the pro-enzyme region precludes expression of an active enzyme by F32-ART5 parasites. Gene inactivation has shown to induce a transient reduction of hemoglobin degradation compensated by expression of other members of the cysteine proteinases family, with minimal impact on growth rate^{57,58}. However, *falcipain 2a* is the only gene from the list of seven affected loci that has been associated with the *in vitro* response to artemisinin. Indeed, it has been convincingly shown that inhibition of falcipain2a-dependent hemoglobin digestion by specific inhibitors or by gene inactivation reduced parasite susceptibility to artemisinins¹⁹. Moreover, ring stages that do not massively digest hemoglobin display a reduced susceptibility to artemisinins⁵⁹.

PF3D7_1302100 (PF13_0011), is a one-exon gene that codes for the gamete antigen 27/25 (Pfg27) produced at the onset of gametocytogenesis. The gene is specific to *P. falciparum* and its close relatives such as *P. reichenowi*. This is an abundant, dimeric phosphorylated cytoplasmic protein that binds RNA. The various KO lines generated display conflicting phenotypes some being deficient in gametocytogenesis⁶⁰, while other Pfg27-defective lines undergo unimpaired gametocytogenesis up to stage V, mature gametocytes although absence of Pfg27 is associated with abnormalities in intracellular architecture of gametocytes⁶¹. The crystal structure shows that the protein forms a dimer, displays a particular RNA binding fold and possesses two Pro-X-X-Pro motifs (known ligands for various domains, including SH3 modules), which combine to form a receptacle for SH3 modules⁶². The P201T mutation is located in the C-terminal ProX-X-Pro motif and predicted to alter the spatial structure of the interaction domain and thus have functional consequences.

PF3D7_1459600 (PF14_0569), is a two-exon gene that codes for a 806 residue-long, conserved protein of unknown function. The *P. yoelii* ortholog has been annotated as the CAAT-box DNA binding subunit B. Close orthologs can be found only among the *Plasmodium* species. Proteomics data indicate that the protein is present in asexual (trophozoites, schizonts, merozoites and rings) and sexual (gametocytes) blood stages of *P. falciparum*. A predicted approx. 130 aa-long Interpro domain suggests presence of an N-terminal multi-helical, alpha-alpha 2-layered structural VHS fold, possibly involved in intracellular membrane trafficking. The rest of the coding sequence carries no specific domain signature. The S299T mutation is located within this "unknown" region.

PF3D7_1464500 (PF14_0603), is a five-exon gene that codes for a 3251 residue-long protein of unknown function, with 4 predicted trans-membrane domains, but otherwise no specific domain signature. Apart from proteomics data indicating its expression and phosphorylation in schizonts, with possible expression in gametocytes and sporozoites as well, little is known about its putative function. The N1629S mutation is located in the middle of the protein, with unpredictable phenotypic impact.

Extended Data Table 4 | Geographic origin and year of collection of archived blood samples studied for K13-propeller polymorphism

Region	Province	Year of collection					Total	
		2001-2002	2003-2004	2005-2006	2007-2008	2009-2010		2011-2012
Western Cambodia	Battambang	64	0	0	0	0	71	135
	Pailin	40	43	46	95	66	84	374
	Pursat	0	10	0	0	43	19	72
Southern Cambodia	Kampot	0	0	0	0	0	12	12
	Kampong Som	0	0	0	0	0	7	7
Northern Cambodia	Oddar Meanchey	0	0	0	0	0	33	33
	Preah Vihear	27	27	25	24	0	19	122
Eastern Cambodia	Kratie	15	0	0	0	0	17	32
	Monduliri	0	0	0	0	0	3	3
	Ratanakiri	56	30	22	0	8	35	151
Total		202	110	93	119	117	300	941

Extended Data Table 5 | Polymorphisms observed in the K13-propeller in Cambodian *P. falciparum* isolates collected in 2001–2012 and in The Gambia (ref. 42)

Codon Position	Amino Acid reference	Nucleotide reference	Amino Acid mutation	Nucleotide mutation
449	G	ggt	A	gCt
458	N	aat	Y	Tat
474	T	aca	I	aTa
476*	M	atg	I	atA
481	A	gct	V	gTt
493	Y	tac	H	Cac
508	T	act	N	aAt
527	P	cct	T	Act
533	G	ggt	S	Agt
537	N	aat	I	aTt
539	R	aga	T	aCa
543	I	att	T	aCt
553	P	ccg	L	cTg
561	R	cgt	H	cAt
568	V	gtg	G	gGg
574	P	cct	L	cTt
580	C	tgt	Y	tAt
584	D	gat	V	gTt
612**	E	gaa	D	gaT
623	S	agt	C	Tgt

* Observed in F32-ART5, not observed in Cambodia

** Reported in The Gambia⁴², not observed in Cambodia

Extended Data Table 6 | Association between polymorphisms observed in the K13-propeller and KH subpopulations (ref. 15) in 150 *P. falciparum* isolates collected in 2009–2010 in Pursat ($n = 103$) and Ratanakiri ($n = 47$) provinces, Cambodia

KH group	Province	Mutations in the K13-propeller				Total
		Wildtype	C580Y	R539T	Y493H	
KH1	Pursat	7	0	0	2	9
	Ratanakiri	46	0	0	0	46
KH2	Pursat	0	25	0	1	26
	Ratanakiri	0	0	0	0	0
KH3	Pursat	3	7	4	0	14
	Ratanakiri	0	0	0	0	0
KH4	Pursat	0	0	0	12	12
	Ratanakiri	0	0	0	0	0
KHA	Pursat	15	19	2	6	42
	Ratanakiri	1	0	0	0	1
Total		72	51	6	21	150