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Population structure of multidrug-resistant *Mycobacterium tuberculosis* clinical isolates in Colombia

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22 **Abstract**

23 Emergence of multidrug-resistant (MDR) *Mycobacterium tuberculosis* complex (MTBC)
24 isolates is a major public health problem that threatens progress made in tuberculosis (TB) care and
25 control worldwide. In Colombia, the prevalence of MDR tuberculosis (MDR-TB) has increased slowly
26 but steadily since 2001. However, the population structure of the MDR-TB strains circulating in
27 Colombia is sparsely known. In this work, 203 MDR isolates isolated in 2012-2013 were collected, and
28 characterized by spoligotyping, followed by 24-loci MIRU-VNTR (data available for 190 isolates). The
29 most prevalent genotypes corresponded to SIT42/LAM9 (12.81%), SIT62/H1 (10.34%), and
30 SIT190/Beijing (10.34%). A fine analysis showed that although the MDR strains came from 29 of the
31 33 departments of Colombia, the distribution of these main lineages was not at random and depended
32 on the city of isolation (p -value <0.000001). Both LAM and Beijing lineage strains were significantly
33 associated with MDR-TB (p -value <0.0001): LAM lineage was associated with 2 patterns of MDR,
34 namely combined resistance to INH + Rifampin (HR), and to SHRE (Streptomycin + INH + Rifampin +
35 Ethambutol), while the Beijing lineage strains were essentially associated with MDR (SHRE).
36 Interestingly, distribution of genotypic lineages in function of drug resistance information (e.g.
37 pansusceptible vs. MDR) was different in our setting as compared to other countries in Latin America.
38 However, MIRU-VNTR patterns were unique for all strains, an observation that did not support active
39 transmission of circulating MDR clones.

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41 Key words: Multidrug-resistant, molecular epidemiology, tuberculosis, Colombia.

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55 1. Introduction

56 In 2018, according to the World Health Organization (WHO), there were approximately about
57 half a million (range: 417 000–556 000) new cases of rifampicin-resistant TB (of which 78% had
58 multidrug-resistant TB [1]. These forms of TB are becoming a worldwide public health problem that
59 requires fast and effective action for its control.

60 Colombia with a population near to 50 million inhabitants represents a developing country
61 where conditions of poverty and inequity are at the same time, an obstacle, and a challenge to the TB
62 control program. In Colombia, the incidence of TB is 26.9 cases per 100,000 inhabitants and for the
63 year 2018, Public Health Surveillance System (Sivigila) reported 14,338 TB cases, of which 428 were
64 drug-resistant, of these 211 were MDR and RR (rifampicin-resistant) [2,3]. At the national level, the
65 prevalence of MDR-TB was 2.38% between the years 2004-2005 in untreated patients. Since then,
66 there has been an increase in the number of cases of MDR-TB: with 601 cases being diagnosed from
67 2010 to 2013 [2]. However, considering WHO estimates of 580 new cases of MDR-TB per year [1], the
68 cases effectively diagnosed are probably an underrepresentation of the real situation. Furthermore,
69 monitoring of MDR-TB treatment in Colombia for the year 2009 revealed a relatively lower treatment
70 success rate of 43% [2], which is a matter of concern for TB control in the country. During the period
71 between 2009 and 2019, an increase in drug-resistant TB was observed (from 7 cases in 2009 to 427
72 cases in 2019), thanks to a better access to molecular drug detection and sensitivity tests, and
73 improvement in the notification of these cases [3].

74 In such a context, improved epidemiologic surveillance today makes use of molecular markers
75 such as IS6110-RFLP, genetic polymorphism based on the variability of the Direct Repeat (DR) locus
76 (spoligotyping), and the mycobacterial interspersed repetitive-unit-variable-number tandem-repeats
77 (MIRU-VNTRs) in order to identify the circulating strains [4,5]. As reviewed recently [6–8],
78 spoligotyping in conjunction with other markers such as Large Sequence Polymorphisms (LSPs) and
79 whole genome sequence techniques has allowed to reveal the worldwide population structure of *M.*
80 *tuberculosis*. Briefly, the global population structure of the *M. tuberculosis* has been composed by
81 seven LSP-based lineages [9–11] or around 12 spoligotyping based lineages [12,13]. The seven LSP-
82 based lineages are known as Indo-Oceanic (L1), East Asian (L2), Indian-East African (L3), Euro-
83 American (L4), West African I & II (L5, L6) and Ethiopian (L7) [6-8]. In SITVIT databases [12,13], these
84 lineages correspond to the following spoligotyping-defined lineages: L1 corresponds to East-African-
85 Indian (EAI); L2 corresponds to Beijing; L3 corresponds to Central-Asian (CAS); L4 is split in at least
86 five lineages (Haarlem or H, X, Latin-American-Mediterranean or LAM, S, and T); L5 and L6
87 correspond to AFRI or *M. africanum*; while L7 has been recently defined in the database and
88 corresponds to Ethiopian lineage. More recently, a new lineage named L8 – seemingly restricted to
89 the African Great Lakes region – was described; phylogenomic analysis showed that L8 is a sister
90 clade to the MTBC lineages [14]. Furthermore, another new lineage, L9, was recently discovered [15].

91 Interestingly, study of global phylogeography of tubercle bacilli not only suggested a co-
92 evolution of MTBC lineages with the human host [11, 14] but further suggested that different *M.*

93 *tuberculosis* lineages could have different predispositions for the acquisition of multidrug resistance;
94 e.g., L2/Beijing strains acquired drug resistances in vitro more rapidly than L4/ Euro-American strains
95 [16]. However, studies in human population showed that diverse genotypes have similar frequencies
96 of acquisition of drug-resistant TB [17,18].

97 Hence, a better knowledge of the genetic characteristics of circulating MTBC strains seems a
98 valid prerequisite not only to better comprehend the prevailing epidemiological snapshot, but also to
99 implement a rational and successful therapy. In this regard, previous studies on MTBC population
100 structure in Colombia showed the high proportion of LAM and Haarlem lineages [19–21] a result that
101 was as expected due to the preponderance of the L4/Euro-American strains in the Americas.
102 Nevertheless, studies analyzing MDR-TB strains in Valle del Cauca, the second department in
103 caseloads in Colombia, indicated that not only LAM but also the Beijing genotype were significantly
104 associated with MDR and XDR-TB isolates [22–25]. Consequently, the present study aimed to provide
105 with an up-to-date inventory of the genetic diversity of MDR-TB isolates circulating in Colombia by
106 studying all such isolates obtained between 2012 and 2013. The spoligotyping-based lineages were
107 further compared to circulating MTBC genotypes not only in Colombia but also in neighboring Latin
108 American countries using an international genotyping database.

109 **2. Materials and Methods**

110 **2.1 Ethical considerations**

111 This study provides with data on retrospective genotyping of a total of 203 MDR-TB isolates
112 collected between the years 2012-2013 from 29 of the 33 territorial entities of Colombia. Considering
113 the total of 233 cases of MDR TB reported in Colombia for this period [2] (n=97 in 2012 and n=136 in
114 2013), the 203 isolates analyzed correspond to 77% of all cases of MDR TB in Colombia. The strains
115 in question were processed by the National Network of Laboratories under their routine activity on TB
116 diagnosis and sent to the National Reference Laboratory at the National Institute of Health in Bogotá,
117 for confirmation and evaluation of drug resistance. Note that the results of the second-line drug
118 susceptibility tests were from previously informed providers and territorial health entities, the latter
119 being responsible for treatment under established national TB guidelines. Retrospective genotyping of
120 MTBC strains met with the standards set out in resolution 008430 of 1993 by the Ministry of Social
121 Protection, Colombia, as follows. Briefly, (i) all sampling and testing were made as part of diagnosing
122 the disease and drug resistance for patient benefit, (ii) retrospective genotyping did neither require
123 additional sampling / new intervention, nor any risk for the evaluated cases, and (iii) all genotyping
124 data were anonymized prior to reporting to the TB genotyping database curator for an anonymous
125 comparison focusing on genotypic and phylogenetic analysis of MTBC genetic diversity.

126 **2.2 Bacterial isolates, genotyping, and database comparison**

127 All *Mycobacterium tuberculosis* complex (MTBC) isolates showing resistance either to
128 isoniazid and/or rifampicin by molecular or conventional tests by the National Network of Laboratories,
129 in Colombia by the National Network of Laboratories were sent for confirmation and evaluation of drug

130 resistance to the National Reference Laboratory at the National Institute of Health. The strains were
131 cultured in Lowenstein-Jensen medium and MGIT liquid medium (BD Diagnostics, Sparks MD, USA),
132 subjected to routine species identification [26], followed by first- and second-line drug susceptibility
133 testing (DST) using Bactec™ MGIT™ 960 methodology according to the manufacturer's
134 recommendations with following drug concentrations: First-line: streptomycin (S), 1µg/ml and 4µg/ml;
135 isoniazid (H), 0,1µg/ml and 0,4µg/ml; rifampicin (R), 1µg/ml; ethambutol (E), 5µg/ml and 7,5µg/ml; and
136 Second-line: kanamycin (Km), 2,5 µg/ml; amikacin (Am), 1µg/ml; capreomycin (Cm), 2,5 µg/ml;
137 ofloxacin (Of), 2 µg/ml. After DST screening, a total of 203 MTBC isolates (1 strain per patient)
138 isolated between the years 2012-2013 from 29 of the 33 territorial entities of Colombia, were retained
139 for this study.

140 The isolates were spoligotyped as reported [4], the results obtained were converted into binary
141 and octal codes and analyzed using the SITVIT2 proprietary database of the Pasteur Institute of
142 Guadeloupe [13,27] which is an updated version of previously released SpoIDB4 and SITVITWEB
143 databases [12]. At the time of this study, SITVIT2 contained a total of 111,635 MTB clinical isolates
144 from 169 countries of patient origin. In this database, Spoligotype International Type (SIT) designates
145 identical patterns shared by two or more isolates, whereas “orphan” designates unique patterns
146 reported for a single isolate. Major phylogenetic lineages were assigned according to spoligotype
147 signatures using SITVIT2 database and for the 7 major MTBC genetic lineages we used the CBN
148 method from TB-Lineage (http://tbinsight.cs.rpi.edu/run_tb_lineage.html) [28]. The lineage distribution
149 in cities or departments of Colombia was compared with those from other cities of Colombia, as well
150 as other countries in Latin America (Mexico, Venezuela, Brazil and Peru), for which data are available
151 in the SITVIT2 database. Lastly, the worldwide distribution of clusters containing 5 or more isolates in
152 this study, and their worldwide distribution was studied both at the country (2 letter country codes
153 according to http://en.wikipedia.org/wiki/ISO_3166-1_alpha-2), as well as macro-geographical level
154 (United Nations subregions at <http://unstats.un.org/unsd/methods/m49/m49regin.htm>). 24-MIRU-
155 VNTR typing [5] was performed using PCR followed by agarose gel electrophoresis, PCR products
156 were analyzed in a photo documenter (Bio Imagen System, ChemiGenius model), and the molecular
157 weight of each fragment has been automatically determined using GeneSnap® software version 6.07
158 GeneTools.

159 **2.3 Phylogenetical and statistical analyses**

160 To establish the relationships between different isolates of MDR-TB we calculated the
161 similarity tree using the unweight pair group method with arithmetic averages (UPGMA) using the
162 MIRU-VNTRplus web application (<http://www.miru-vntrplus.org/MIRU/index.faces>) [29]. A cluster was
163 defined as two or more *M. tuberculosis* isolates with identical patterns (note that the term “cluster” is
164 used here for spoligotyping data solely for convenience and does not imply recent transmission). The
165 evolutionary relationships among all the observed spoligotypes and MIRU-VNTR patterns were
166 obtained by drawing minimum spanning trees (MSTs) using MLVA Compare software from Geno
167 Screen and Ridom Bioinformatics and BioNumerics version 6.6 ([https://www.applied-
168 maths.com/bionumerics](https://www.applied-maths.com/bionumerics)). MSTs are undirected graphs in which all samples are connected with the

169 fewest possible connections between nearest neighbors. Finally, statistical analysis was performed
170 using the R software. Chi-squares and P-values were calculated with Pearson's Chi-squared test. A p-
171 value between 0.05 and 0.07 was considered marginally significant. A p-value<0.05 was considered
172 statistically significant. The Hunter-Gaston Discriminatory Index (HGDI) was used to evaluate the
173 discriminatory power of the spoligotyping method.

174 **3. Results**

175 **3.1 Characteristics of the population studied**

176 We analyzed two hundred and three MDR isolates that were collected from 29 of 33 territorial
177 entities of Colombia during the years 2012-2013. Most MDR-TB cases occurred in the department of
178 Antioquia, mainly due to the cases brought by the capital city Medellin, which is the city with the
179 highest MDR-TB isolates (29.56%, n=60) (Table S1). The second is Valle del Cauca department
180 (23.16%), n=47), whose cases are mostly provided by Buenaventura (14.29%, n=29), which is a port
181 city boarding the Pacific Ocean, and the capital Cali (8.87%, n=18). The third department that has
182 more cases of MDR-TB is Atlántico, whose cases are concentrated in the capital Barranquilla, a port
183 boarding the Caribbean Sea. The fourth department is Norte de Santander whose cases are in the
184 capital Cucuta (3.45%, n=7) which is a border town with Venezuela. Medellin and Cali are cities with
185 more than two million people, meanwhile Buenaventura has 400.000 people and Cucuta has 650.000
186 people (<http://www.dane.gov.co/index.php/estadisticas-por-tema/demografia-y-poblacion>). The global
187 Male/Female sex ratio was 129/74, i.e. 1.74 and the mean age was 39 year, with the interquartile
188 range between 26 to 53 years of age (Table S2).

189 **3.2 MDR-TB Population Structure**

190 To determine the population structure of the 203 MDR-TB strains, we carried out the spoligotyping
191 technique. We found 52 SITs, twenty-seven SITs were clustered (51.9%) meanwhile twenty-five SITs
192 were unique (48.1%). A total of 6 SITs were recently created and 4 of them were exclusively found in
193 Colombia (Table 1). Among these newly created SITs, one pattern belonged to an "unknown"
194 signature (SIT4100), another to X lineage (SIT4105), while the remaining 4 patterns (SIT4101 to 4104)
195 belonged to LAM. Lastly, the description of clusters containing 5 or more isolates and their worldwide
196 distribution according to the SITVIT database is shown in Table 2. The most dominant spoligotype
197 family in the MDR cases was Latin American and Mediterranean (SIT42/LAM9, 12.81%, n=26),
198 Haarlem (SIT62/H1, 10.34%, n= 21), Beijing (SIT190/Beijing, 10.34%, n=21), followed by the ill-
199 defined T (SIT53/T1, 4.43%, n=9). Fourteen isolates were orphans. A dendrogram was constructed
200 based on spoligotyping results (Figure S1); according to this tree 163/203 isolates were grouped into
201 27 clusters, containing from 2 to 26 isolates; the most dominant clusters were represented by the most
202 dominant SITs, as mentioned previously. In this study, the discriminatory power of the spoligotyping
203 method was 0.9464 measured by the Hunter-Gaston Discriminatory Index (HGDI).

204 24-loci MIRU-VNTR analysis showed an absence of clustering; furthermore, the patterns observed did
205 not match with the patterns previously submitted to the SITVIT database, hence classified as orphans
206 (Table S4). However, despite the absence of clustering, the MST tree (n=190 isolates) obtained with

207 the combined information on 24-loci MIRU-VNTR and spoligotyping based lineages, showed that the
 208 strains tended to group together in function of their spoligotyping pattern (Figure S2). For example, the
 209 branches corresponding to the LAM, Haarlem, and Beijing lineages were clearly well defined. The
 210 most represented lineage in the tree was LAM with 40% of isolates (n=76/190), followed by Haarlem
 211 22,1% (n=42/190) and Beijing 11% (n=21/190). Lineages S, X and T were less well defined in the tree
 212 and corresponded to 22% of the isolates. The other well-defined branch in the tree corresponded to
 213 unknown lineage isolates (n=10/19; 0,05%;) though most of them belonged to the SIT881. In the
 214 SITVIT2 database, SIT881 was present in Venezuela, United States, Italy, Spain and Colombia;
 215 nevertheless, Colombia contributed with 72% (n=18/25) of such isolates. Because of the high
 216 variability in the MIRU-VNTR patterns, it was difficult to find an allelic marker for the branches
 217 observed; and in this regard, the loci with the highest polymorphism were QUB 26 (0.87), MIRU10
 218 (0.82) and QUB11b (0.81). Further analyses would be needed to better delineate sublineages from
 219 genotyping data as performed in another study [30].

220 3.3 Lineage distribution analysis

221 We determined the distribution of the eight lineages/sublineages found in this study: Beijing,
 222 EAI, Haarlem, LAM, Manu, S, T and X. LAM was the most dominant lineage with 36.45% (n=74),
 223 followed by Haarlem with 23.15% (n=47), Beijing and T, each one with 10.34% (n=21), X with 7.39%
 224 (n=15), S with 2.46% (n=5) and finally EAI with 0.49% (n=1). Remarkable was the presence of 17
 225 isolates with unknown lineage (8.37%) (Table S3, A). The difference of lineage/sublineages
 226 distribution between the two years (2012 and 2013) was “marginally” significant (p-value=0.0504)
 227 (Table S3, B). The difference between gender of patients vs. lineage was not statistically significant (p-
 228 value=0.88). When comparing distribution of main lineages/sublineages (Beijing, Haarlem, LAM and
 229 T) in Medellin (n=68 isolates), Buenaventura (n=29 isolates), Cali (n=18 isolates), and other cities
 230 grouped together (n=96 isolates), we noted a statistically significant difference (p-value<0.000001)
 231 (Table 3). Beijing lineage was more frequent in Buenaventura, Haarlem lineage was more frequent in
 232 Medellin, and LAM was more frequent in Cali as well as the group of other cities. A detailed map of
 233 Colombia showing the phylogeographical distribution of MTB lineages/sublineages is illustrated in
 234 **figure 1**.

235 The department with the highest diversity of lineages/sublineages is Valle del Cauca (where
 236 Buenaventura and Cali are located; (n=16), followed by Antioquia (with Medellin being the capital;
 237 n=11), Atlántico (n=8), and Bogotá and Norte de Santander (n=6 each). Moreover, the distribution of
 238 the lineages/sublineages is different in each region, as mentioned above. To sum up, a statistically
 239 significant difference was noted when comparing MDR (HR), MDR (SHR), and MDR (SHRE) groups
 240 vs. lineages; note however that not enough data were available for MDR (HRE) group. Last but not
 241 least, 20/21 of strains belonging to Beijing lineage were associated with MDR (SHRE), while a great
 242 proportion of LAM lineage strains was associated with MDR (HR) (p-value<0.0001) (Table 3).

243 Comparing the distribution of MDR-TB lineages/sublineages obtained in this study (Colombia)
 244 with the data available in SITVIT2 for other Latin American countries such as Mexico, Venezuela, Peru
 245 and Brazil, we noticed that the most dominant lineage among the MDR-TB isolates varied according to

246 the countries studied – thus it was LAM in Colombia, Venezuela, Brazil and Peru, whereas the
247 sublineage T was the most dominant lineage in Mexico (45.30%, n=82) (**Figure 2**). The Manu sub
248 lineage was only present in Colombia and Brazil. In a nutshell, each country had its own characteristic
249 distribution, for example, the most prominent lineages/sublineages were LAM, Haarlem, Beijing and T
250 in Colombia, as opposed to T, X and LAM in Mexico.

251 **3.4 Evolutionary relationships**

252 Minimum spanning trees (MSTs) shown in Figure 3 were based on all spoligotypes of this
253 study (n=203 isolates). **Figure 3A** illustrates evolutionary relationships of spoligotypes in function of
254 MTBC lineages. One may notice that the most visible lineages/sublineages were LAM, Haarlem,
255 Beijing, and T. Only 2 isolates belonged to Manu sublineage, and only one strain belonged to EAI
256 (East-African-Indian). The most visible SITs were SIT42/LAM9 (n=26 isolates), SIT62/H1 (n=21
257 isolates), SIT190/Beijing (n=21 isolates), SIT53/T1 (n=9 isolates), SIT50/H3 (n=8 isolates), SIT217/X1
258 (n=8 isolates), and SIT881/Unknown (n=8 isolates). Some newly created SITs (such as SITs 4101,
259 4102, and 4104) as well as some orphan spoligotypes (Or06, Or10, and Or12), and SIT2648
260 belonging to LAM sublineage appeared at a terminal position in the tree, forming a sub-branch rooted
261 by SIT4101. These spoligotypes may be specific to Colombia, and may constitute a subgroup of LAM
262 sublineage marked by the absence of spacers 5 and 15. **Figure 3B** is the same tree as the precedent,
263 but it was drawn in function of information on first-line MDR-TB drugs. One may notice that
264 SIT190/Beijing, appearing at the bottom of the MST, was highly associated with MDR (SHRE) group.
265 SIT42/LAM9 was mostly associated with both MDR (HR) and MDR (SHRE). Furthermore, a high
266 proportion of MDR (SHRE) was also visible among isolates belonging to SIT62/H1.

267 The MST analysis based on 24-loci MIRU-VNTR data (n=190 isolates) showed that all
268 patterns were unique (as confirmed by comparison with SITVIT database). However, despite their
269 uniqueness and diversity, we noticed that isolates were rather well organized in function of
270 spoligotyping based lineages, as mentioned before (**Figure 4**). The UPGMA tree drawn in **Figure S2**
271 showed that strains belonging to SIT42/LAM9 and SIT53/T1 were particularly scattered in different
272 branches. This observation corroborated the fact that several sublineages could be discovered among
273 strains sharing a same spoligotyping pattern.

274

275 **4. Discussion**

276 The present work characterizes the genotypes of the 203 MDR-TB isolates recollected during
277 2012-2013 from 29 of 33 territorial entities of Colombia. The predominant genotypes within this pool of
278 MDR-TB isolates belonged to the most predominant lineages present in Colombia reported previously
279 [19–21], i.e. LAM and Haarlem sublineages represented by the SIT42/LAM9 (12.81%,) and SIT62/H1
280 (10.34%) respectively. However, we must highlight the presence of a specific Beijing genotype
281 (SIT190/Beijing) as predominant genotype in MDR-TB isolates, mainly isolated in the seaport of
282 Buenaventura (Valle del Cauca department) (Table 3). Although the current data might not be
283 sufficient to make year to year assessment, prevalence of SIT190/Beijing strains increased from 7

284 isolates in 2012 to 14 in 2013. This trend was already observed previously in a cohort of patients from
285 1999 to 2012 regarding MDR-TB isolates associated with the Beijing lineage [21]. Indeed, the high
286 prevalence of Beijing genotype in the city of Buenaventura has been reported since 1998 [22], with
287 subsequent emergence of a so-called rare "Beijing-like" genotype SIT190 [23], which continues to
288 dominate the MDR-TB cases in Buenaventura [24,25]. Apart from human migration, this may partly be
289 due to deficiencies in the monitoring and treatment of patients by the precarious health system in
290 Colombia. Indeed, despite the recommendations made by the public-private organizations from Valle
291 del Cauca, interested in knowing the MDR-TB situation in this department, much remains to be done
292 to precisely know the proportion of primary vs. previously treated MDR cases, a finding that will have
293 significant impact on TB control and treatment outcome [31].

294 We observed a strong association between Beijing genotype and resistance to isoniazid,
295 rifampicin, ethambutol and streptomycin (20/21 strains, p-value < 0.0001). Several studies have
296 shown that *M. tuberculosis* strains belonging to the L2/East-Asian lineage, to which Beijing genotype
297 belongs, have strong association with drug-resistance and increase virulence [32–34]. A study showed
298 that particular strains of the Beijing genotype acquired drug resistances in vitro more rapidly than L4/
299 Euro-American strains [16], but this observation has not been confirmed in real settings. The
300 association of Beijing strains with MDR-TB can also differ depending on countries in Latin America
301 (Fig. 2) [35] and elsewhere [13]. In this regard, as compared data shown for Latin American countries
302 illustrated in Fig. 2 (Colombia, Peru, Brazil, and Venezuela), a recent study from Peru highlighted the
303 fact that LAM lineage strains were more likely to be drug resistant as compared to Beijing strains [36],
304 while T lineage strains were associated with MDR-TB in Mexico [37]. In conclusion, despite
305 geographic variability observed for association of MDR-TB and Beijing genotype at a global scale [as
306 reviewed recently; [13]], our results do provide with clinical and epidemiological evidence that
307 L2/Beijing genotype strains, and in particular an emerging SIT190/Beijing variant, show an increased
308 capacity to be transmitted and be associated with drug resistance in Colombia [38,39].

309 Considering the prevailing worldwide occurrence of Beijing strains and their dominance in
310 certain countries [40,41], one must be extremely careful with the subtle change in the genetic diversity
311 landscape and drug-resistance characteristics of prevailing *M. tuberculosis* strains in Colombia.
312 Remarkably, a high diversity of lineages is found in Valle del Cauca (n=13) with predominance of the
313 Beijing lineage (see results and Fig. 1) fueled by the seaport of Buenaventura on the Pacific Ocean. In
314 contrast, low diversity and different lineages were observed in the department of Atlántico (n=8; Fig. 1)
315 as compared to Valle del Cauca even though the provider city of Barranquilla is also a port on the
316 Caribbean Sea. Thus, despite both Buenaventura and Barranquilla being port cities, the diversity of
317 lineages and MDR-TB may be explained by the magnitude of the commercial movements in the port
318 of Buenaventura, which moves 33% of exports and 49.6% of Colombian imports
319 (<http://colombiatrader.com.co/>). The high prevalence of the Beijing lineage in Buenaventura,
320 specifically genotype SIT 190/Beijing variant, suggests that it came from China; in fact, the SIT190 is
321 present in China and is the country that contributes the most to this genotype worldwide (24%)
322 (<http://www.pasteur-guadeloupe.fr:8081/SITVIT2/>).

323 According to the 2011 census conducted by the “Departamento Administrativo Nacional de
324 Estadística” (www.dane.gov.co/), Buenaventura with primarily an Afro-Colombian population has a
325 high proportion of people with unmet basic needs (34.52%), while Barranquilla with as little as 13.2%
326 of Afro-Colombians has a significantly lower proportion (17.70%) of people with unmet basic needs. It
327 is therefore tempting to speculate that a high proportion of people with a specific ethnicity living in
328 poverty could be one of the factors favoring the emergence of drug-resistant Beijing genotype
329 variant(s) in Buenaventura. Additionally, MIRU-typing showed that all 24-loci MIRU-VNTR patterns
330 were unique (or orphans as compared to SITVIT database; Table S4) – an observation that did not
331 support active transmission of circulating MDR clones. MDR thus most probably emerged among
332 unrelated cases due to lack of compliance.

333 In concordance with prevailing MTB lineages present among pulmonary TB patients in
334 Colombia [20], Antioquia with the capital city of Medellin was the second department with a high
335 genetic diversity (n=11) in our study, with Haarlem being the most predominant lineage (p-
336 value<0.0001). Strangely enough, Bogotá – the capital city of Colombia, with nearly eight million
337 population, was characterized by least genetic diversity. This result should be re-explored in future
338 investigations to verify if it could be linked to access to good healthcare and better life conditions or
339 simply an underreporting of TB cases. Finally, this study highlighted the presence of new spoligotypes
340 (defined by the absence of spacers 5 and 15), which may constitute a new subgroup of LAM lineages.
341 However further investigations would be necessary to know if these new patterns are specific to
342 Colombia possibly due to specific host-pathogen interactions, or due to a simple coincidence. To sum
343 up, our study brings a deep overview regarding MDR-TB situation in Colombia as compared to other
344 Latin American countries and argues in favor of better strategies and management for control and
345 prevention of TB in Colombia. A limitation of our study is the fact that Whole Genome Sequencing
346 (WGS) analysis was not performed. Future studies should target WGS data to know if active
347 transmission of MDR-TB clones is ongoing and at what extent.

348

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352

353 **Author contribution**

354 MIM designed the study. JGRC, CLL, LACH and JG conducted the experiments. CLL
355 collected the samples and prepared patient’s data. JGRC, CLL, LACH, DC, NR and MIM analyzed
356 and interpreted the data. JGRC, DC and NR collectively wrote the text, and NR edited the manuscript.
357 All authors reviewed and approved the final manuscript.

358

359 **Competing interests**

360 The authors declare there are no competing interests.

361

362 **References**

- 363 [1] World Health Organization. Global tuberculosis report 2019. World Health Organization.
364 <https://apps.who.int/iris/handle/10665/329368>.
- 365 [2] Instituto Nacional de Salud. Informe del Evento Tuberculosis Farmacorresistente, periodo
366 epidemiológico 12 del año 2013. Bogotá D.C. 2013. [https://www.ins.gov.co/buscador-](https://www.ins.gov.co/buscador-eventos/Lineamientos/PRO_Tuberculosis.pdf)
367 [eventos/Lineamientos/PRO_Tuberculosis.pdf](https://www.ins.gov.co/buscador-eventos/Lineamientos/PRO_Tuberculosis.pdf)
- 368 [3] Cruz-Martinez OA. Comportamiento de la tuberculosis y avances en la implementación del
369 plan estratégico hacia el fin de la tuberculosis en Colombia 2016-2025. Bogotá D.C. 2020.
370 <https://www.risaralda.gov.co/descargar.php?idFile=14217>
- 371 [4] Kamerbeek J, Schouls L, Kolk A, Van Agterveld M, Van Soolingen D, Kuijper S, et al.
372 Simultaneous detection and strain differentiation of *Mycobacterium tuberculosis* for diagnosis
373 and epidemiology. J Clin Microbiol 1997;35: 907-14. [https://doi.org/10.1128/JCM.35.4.907-](https://doi.org/10.1128/JCM.35.4.907-914.1997)
374 [914.1997](https://doi.org/10.1128/JCM.35.4.907-914.1997).
- 375 [5] Supply P, Allix C, Lesjean S, Cardoso-Oelemann M, Rüsch-Gerdes S, Willery E, et al.
376 Proposal for standardization of optimized mycobacterial interspersed repetitive unit-variable-
377 number tandem repeat typing of *Mycobacterium tuberculosis*. J Clin Microbiol 2006;44:4498-
378 510. <https://doi.org/10.1128/JCM.01392-06>.
- 379 [6] Firdessa R, Berg S, Hailu E, Schelling E, Gumi B, Erenso G, et al. Mycobacterial lineages
380 causing pulmonary and extrapulmonary tuberculosis, Ethiopia. Emerg Infect Dis. 2013;19:460-
381 3. <https://doi.org/10.3201/eid1903.120256>.
- 382 [7] Blouin Y, Hauck Y, Soler C, Fabre M, Vong R, Dehan C, et al. Significance of the identification
383 in the Horn of Africa of an exceptionally deep branching *Mycobacterium tuberculosis* clade.
384 PLoS One. 2012;7(12):e52841. <https://doi.org/10.1371/journal.pone.0052841>.
- 385 [8] Jagielski T, Minias A, van Ingen J, Rastogi N, Brzostek A, Żaczek A, et al. Methodological and
386 clinical aspects of the molecular epidemiology of *Mycobacterium tuberculosis* and other
387 mycobacteria. Clin Microbiol Rev 2016;29:239-90. <https://doi.org/10.1128/CMR.00055-15>.
- 388 [9] Comas I, Coscolla M, Luo T, Borrell S, Holt KE, Kato-Maeda M, et al. Out-of-Africa migration
389 and Neolithic coexpansion of *Mycobacterium tuberculosis* with modern humans. Nat Genet
390 2013;45:1176–82. <https://doi.org/10.1038/ng.2744>.
- 391 [10] Gagneux S. Genetic diversity in *Mycobacterium tuberculosis*. Curr Top Microbiol Immunol
392 2013;374:1–25. https://doi.org/10.1007/82_2013_329.
- 393 [11] Gagneux S, Small PM. Global phylogeography of *Mycobacterium tuberculosis* and implications
394 for tuberculosis product development. Lancet Infect Dis 2007;7:328-37.
395 [https://doi.org/10.1016/S1473-3099\(07\)70108-1](https://doi.org/10.1016/S1473-3099(07)70108-1).
- 396 [12] Brudey K, Driscoll JR, Rigouts L, Prodinger WM, Gori A, Al-Hajj S a, et al. *Mycobacterium*
397 *tuberculosis* complex genetic diversity: mining the fourth international spoligotyping database
398 (SpolDB4) for classification, population genetics and epidemiology. BMC Microbiol 2006;6:23.
399 <https://doi.org/10.1186/1471-2180-6-23>.
- 400 [13] Couvin D, David A, Zozio T, Rastogi N. Macro-geographical specificities of the prevailing
401 tuberculosis epidemic as seen through SITVIT2, an updated version of the *Mycobacterium*
402 *tuberculosis* genotyping database. Infect Genet Evol 2019;72:31-43.
403 <https://doi.org/10.1016/j.meegid.2018.12.030>.
- 404 [14] Ngabonziza JCS, Loiseau C, Marceau M, Jouet A, Menardo F, Tzfadia O, et al. A sister lineage
405 of the *Mycobacterium tuberculosis* complex discovered in the African Great Lakes region. Nat
406 Commun 2020;11:2917 <https://doi.org/10.1038/s41467-020-16626-6>.
- 407 [15] Coscolla M, Brites D, Menardo F, Loiseau C, Darko Otchere I, Asante-Poku A, et al.
408 Phylogenomics of *Mycobacterium africanum* reveals a new lineage and a complex evolutionary
409 history. BioRxiv 2020;17:19. <https://doi.org/10.1101/2020.06.10.141788>. Posted June 10, 2020
- 410 [16] Ford CB, Shah RR, Maeda MK, Gagneux S, Murray MB, Cohen T, et al. *Mycobacterium*

- 411 *tuberculosis* mutation rate estimates from different lineages predict substantial differences in
 412 the emergence of drug-resistant tuberculosis. *Nat Genet* 2013;45:784–90.
 413 <https://doi.org/10.1038/ng.2656>.
- 414 [17] Chisompola NK, Streicher EM, Muchemwa CMK, Warren RM, Sampson SL. Molecular
 415 epidemiology of drug resistant *Mycobacterium tuberculosis* in Africa: A systematic review. *BMC*
 416 *Infect Dis* 2020;20. <https://doi.org/10.1186/s12879-020-05031-5>.
- 417 [18] Oppong YEA, Phelan J, Perdigão J, Machado D, Miranda A, Portugal I, et al. Genome-wide
 418 analysis of *Mycobacterium tuberculosis* polymorphisms reveals lineage-specific associations
 419 with drug resistance. *BMC Genomics* 2019;20. <https://doi.org/10.1186/s12864-019-5615-3>.
- 420 [19] Cerezo I, Jiménez Y, Hernandez J, Zozio T, Murcia MI, Rastogi N. A first insight on the
 421 population structure of *Mycobacterium tuberculosis* complex as studied by spoligotyping and
 422 MIRU-VNTRs in Bogotá, Colombia. *Infect Genet Evol* 2012;12:657–63.
 423 <https://doi.org/10.1016/j.meegid.2011.07.006>.
- 424 [20] Realpe T, Correa N, Roza JC, Ferro BE, Gomez V, Zapata E, et al. Population Structure
 425 among *Mycobacterium tuberculosis* Isolates from Pulmonary Tuberculosis Patients in
 426 Colombia. *PLoS One* 2014;9:e93848. <https://doi.org/10.1371/journal.pone.0093848>.
- 427 [21] Puerto G, Erazo L, Wintaco M, Castro C, Ribon W, Guerrero MI. *Mycobacterium tuberculosis*
 428 Genotypes Determined by Spoligotyping to Be Circulating in Colombia between 1999 and 2012
 429 and Their Possible Associations with Transmission and Susceptibility to First-Line Drugs. *PLoS*
 430 *One* 2015;10:e0124308. <https://doi.org/10.1371/journal.pone.0124308>.
- 431 [22] Laserson KF, Osorio L, Sheppard JD, Hernández H, Benítez AM, Brim S, et al. Clinical and
 432 programmatic mismanagement rather than community outbreak as the cause of chronic, drug-
 433 resistant tuberculosis in Buenaventura, Colombia, 1998. *Int J Tuberc Lung Dis* 2000;4:673–83.
- 434 [23] Murcia MI, Manotas M, Jiménez YJ, Hernández J, Cortés MIC, López LE, et al. First case of
 435 multidrug-resistant tuberculosis caused by a rare “Beijing-like” genotype of *Mycobacterium*
 436 *tuberculosis* in Bogotá, Colombia. *Infect Genet Evol* 2010;10:678–81.
 437 <https://doi.org/10.1016/j.meegid.2010.03.010>.
- 438 [24] Ferro BE, Nieto LM, Roza JC, Forero L, van Soolingen D. Multidrug-resistant *Mycobacterium*
 439 *tuberculosis*, southwestern Colombia. *Emerg Infect Dis* 2011;17:1259–62.
 440 <https://doi.org/10.3201/eid1707.101797>.
- 441 [25] Nieto LM, Ferro BE, Villegas SL, Mehaffy C, Forero L, Moreira C, et al. Characterization of
 442 extensively drug-resistant tuberculosis cases from Valle del Cauca, Colombia. *J Clin Microbiol*
 443 2012;50:4185–7. <https://doi.org/10.1128/JCM.01946-12>.
- 444 [26] Garzón MC, Restrepo G, Llerena C, Orjuela D, Bueno J, Medina ML. Diagnóstico
 445 bacteriológico de tuberculosis y micobacteriosis. Bogotá, Colombia: Instituto Nacional de Salud;
 446 2012. <https://www.ins.gov.co>
- 447 [27] Couvin D, Rastogi N. The establishment of databases on circulating genotypes of
 448 *Mycobacterium tuberculosis* complex and web tools for an effective response to better monitor,
 449 understand and control the tuberculosis epidemic worldwide. *EuroReference*, 2014;12:36-48.
 450 <https://pro.anses.fr/euroreference/Documents/ER12-RESEAU-MycobacteriumEN.pdf>
- 451 [28] Shabbeer A, Cowan LS, Ozcaglar C, Rastogi N, Vandenberg SL, Yener B, et al. TB-Lineage:
 452 An online tool for classification and analysis of strains of *Mycobacterium tuberculosis* complex.
 453 *Infect Genet Evol* 2012;12:789–97. <https://doi.org/10.1016/j.meegid.2012.02.010>.
- 454 [29] Weniger T, Krawczyk J, Supply P, Niemann S, Harmsen D. MIRU-VNTRplus: A web tool for
 455 polyphasic genotyping of *Mycobacterium tuberculosis* complex bacteria. *Nucleic Acids Res*
 456 2010;38(Web Server issue):W326-31. <https://doi.org/10.1093/nar/gkq351>.
- 457 [30] Mokrousov I, Vyazovaya A, Iwamoto T, Skiba Y, Pole I, Zhdanova S, et al. Latin-American-
 458 Mediterranean lineage of *Mycobacterium tuberculosis*: Human traces across pathogen’s
 459 phylogeography. *Mol Phylogenet Evol* 2016;99:133-143.
 460 <https://doi.org/10.1016/j.ympev.2016.03.020>.

- 461 [31] Villegas SL, Ferro BE, Perez-Velez CM, Moreira CA, Forero L, Martínez E, et al. High initial
462 multidrug-resistant tuberculosis rate in Buenaventura, Colombia: A public-private initiative. *Eur*
463 *Respir J* 2012;40:1569-72. <https://doi.org/10.1183/09031936.00018212>.
- 464 [32] Drobniewski F, Balabanova Y, Nikolayevsky V, Ruddy M, Kuznetsov S, Zakharova S, et al.
465 Drug-resistant tuberculosis, clinical virulence, and the dominance of the Beijing strain family in
466 Russia. *J Am Med Assoc* 2005; 293:2726-31. <https://doi.org/10.1001/jama.293.22.2726>.
- 467 [33] Hanekom M, Gey Van Pittius NC, McEvoy C, Victor TC, Van Helden PD, Warren RM.
468 *Mycobacterium tuberculosis* Beijing genotype: A template for success. *Tuberculosis*
469 2011;91:510–23. <https://doi.org/10.1016/j.tube.2011.07.005>.
- 470 [34] de Steenwinkel JEM, ten Kate MT, de Knecht GJ, Kremer K, Aarnoutse RE, Boeree MJ, et al.
471 Drug susceptibility of *Mycobacterium tuberculosis* Beijing genotype and association with MDR
472 TB. *Emerg Infect Dis* 2012;18:660–3. <https://doi.org/10.3201/eid1804.110912>.
- 473 [35] Cerezo-Cortés MI, Rodríguez-Castillo JG, Hernández-Pando R, Murcia MI. Circulation of *M.*
474 *tuberculosis* Beijing genotype in Latin America and the Caribbean. *Pathog Glob Health* 2019;
475 113:336-351. <https://doi.org/10.1080/20477724.2019.1710066>.
- 476 [36] Grandjean L, Iwamoto T, Lithgow A, Gilman RH, Arikawa K, Nakanishi N, et al. The
477 Association between *Mycobacterium tuberculosis* genotype and drug resistance in Peru. *PLoS*
478 *One* 2015; May 18;10(5):e0126271. <https://doi.org/10.1371/journal.pone.0126271>.
- 479 [37] Martinez-Guarneros A, Rastogi N, Couvin D, Escobar-Gutierrez A, Rossi LMG, Vazquez-
480 Chacon CA, et al. Genetic diversity among multidrug-resistant *Mycobacterium tuberculosis*
481 strains in Mexico. *Infect Genet Evol* 2013;14:434-43.
482 <https://doi.org/10.1016/j.meegid.2012.12.024>.
- 483 [38] Rodríguez-Castillo JG, Pino C, Niño LF, Rozo JC, Llerena-Polo C, Parra-López CA, et al.
484 Comparative genomic analysis of *Mycobacterium tuberculosis* Beijing-like strains revealed
485 specific genetic variations associated with virulence and drug resistance. *Infect Genet Evol*
486 2017;54:314–23. <https://doi.org/10.1016/j.meegid.2017.07.022>.
- 487 [39] Ramirez LMN, Ferro BE, Diaz G, Anthony RM, de Beer J, van Soolingen D. Genetic profiling of
488 *Mycobacterium tuberculosis* revealed “modern” Beijing strains linked to MDR-TB from
489 Southwestern Colombia. *PLoS One* 2020; Apr 24;15(4):e0224908.
490 <https://doi.org/10.1371/journal.pone.0224908>.
- 491 [40] Glynn JR, Whiteley J, Bifani PJ, Kremer K, Van Soolingen D. Worldwide occurrence of
492 Beijing/W strains of *Mycobacterium tuberculosis*: A systematic review. *Emerg Infect Dis*
493 2002;8:843–9. <https://doi.org/10.3201/eid0808.020002>.
- 494 [41] Drobniewski F, Balabanova Y, Nikolayevsky V, Ruddy M, Kuznetsov S, Zakharova S, et al.
495 Drug-resistant tuberculosis, clinical virulence, and the dominance of the Beijing strain family in
496 Russia. *Jama* 2005;293:2726–31. <https://doi.org/10.1001/jama.293.22.2726>.

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499 **Legends to Figures**500 **Fig 1. Phylogeographic distribution of MTBC sublineages among MDR-TB strains in Colombia.**

501 The map shows the distribution of the MDR-TB sublineages corresponding to different territorial
502 entities of Colombia. In dark blue, the territorial entities that have the largest number of MDR-TB
503 cases, each one with its corresponding pie representing the proportion of sublineages found in that
504 region. These territories provide 65% of the isolates from this study. The big pie represents all the
505 lineages found among the pool of MDR-TB isolates.

506 **Fig 2. Map representing distribution of pansusceptible and MDR strains of some Latin**
507 **American countries (Mexico, Venezuela, Brazil, Peru, and Colombia) in the international**
508 **database.** This map was obtained from the following link: http://en.wikipedia.org/wiki/Latin_America
509 according to terms of the Creative Commons 3.0 Attribution License
510 (<http://creativecommons.org/licenses/by/3.0/>). Note that for a better visualization, distribution of strains
511 belonging to this study (n=203) is shown under the subheading Colombia.

512 **Fig 3. A minimum spanning tree (MST) illustrating evolutionary relationships between the *M.***
513 ***tuberculosis* spoligotypes of this study (n=203 isolates).** The phylogenetic tree connects each
514 genotype based on degree of changes required to go from one allele to another (the distance numbers
515 are visible on each edge). Solid black line denotes one unique change between two patterns, while
516 solid gray line denotes 2 changes, bold dashed line denotes 3 changes, and thin dotted line
517 represents 4 or more changes. The size of the circle is proportional to the total number of isolates. SIT
518 numbers appear inside nodes. Trees were drawn in function of phylogenetic lineages (A), and in
519 function of information on first line MDR drugs (B).

520 **Fig 4. A minimum spanning tree (MST) illustrating evolutionary relationships between all 24-**
521 **loci MIRU patterns (n=190 isolates).** All circles have the same size because each pattern was
522 unique. Distance between patterns was represented by dashed lines. The tree was drawn in function
523 of the spoligotyping based lineages.

524 **Table 1.** Description of 52 Spoligotype International Types (SITs; n=189 isolates) and corresponding
525 spoligotyping defined lineages/sublineages starting from a total of 203 *M. tuberculosis* strains isolated
526 in Colombia.

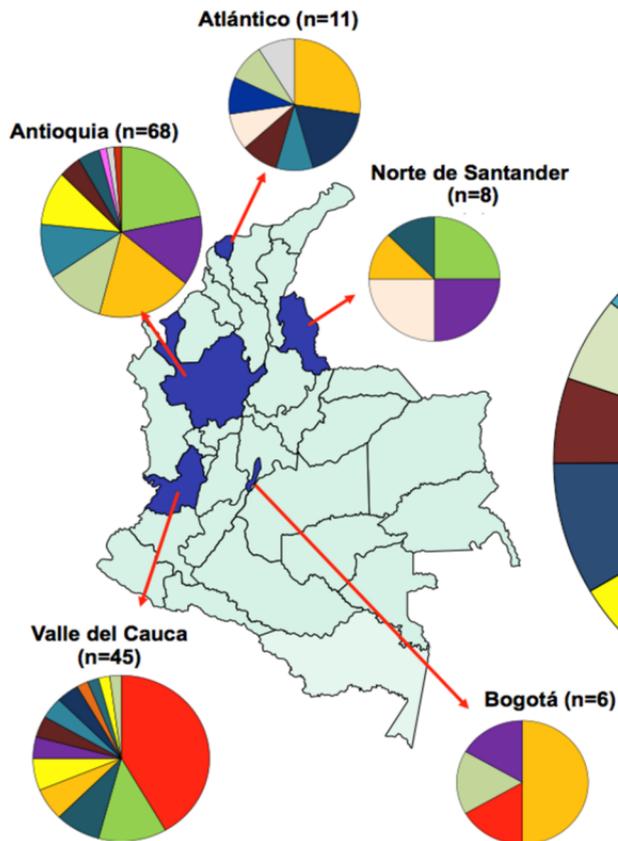
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528 **Table 2.** Description of clusters containing 5 or more isolates in this study, and their worldwide
529 distribution in the SITVIT database

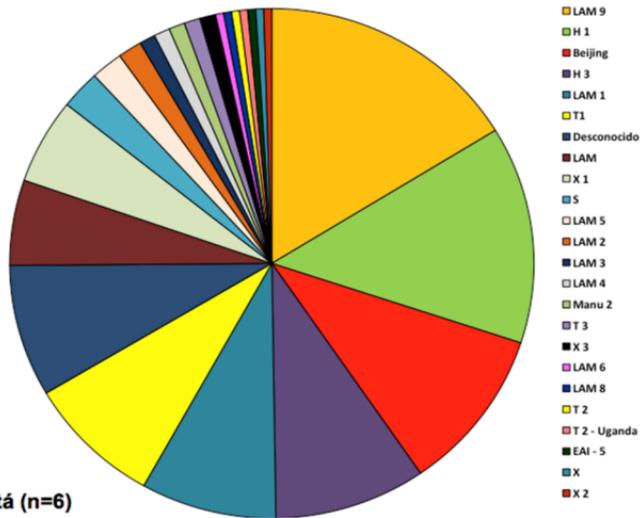
530 **Table 3.** Distribution of MTBC lineages in function of several parameters of the study.

531

532



24 lineages identified (n=203)



Mexico

Lineage	Pan-susceptible (%)	MDR (%)
BOV	5 (3.16)	2 (1.10)
Beijing	2 (1.27)	3 (1.66)
EAI	6 (3.80)	10 (5.52)
Haarlem	16 (10.13)	13 (7.18)
LAM	27 (17.09)	24 (13.26)
Manu	5 (3.16)	0 (0.00)
S	2 (1.27)	9 (4.97)
T	48 (30.38)	82 (45.30)
X	40 (25.32)	24 (13.26)
Unknown	7 (4.43)	14 (7.73)

*P-value<0.005

Venezuela

Lineage	Pan-susceptible (%)	MDR (%)
Beijing	0 (0.00)	2 (3.64)
Haarlem	3 (3.95)	3 (5.45)
LAM	61 (80.26)	41 (74.55)
Manu	1 (1.32)	0 (0.00)
S	1 (1.32)	1 (1.82)
T	7 (9.21)	8 (14.55)
Unknown	3 (3.95)	0 (0.00)

*P-value>0.3

Colombia

Lineage	Pan-susceptible (%)	MDR (%)	This study (%)
Beijing	1 (0.28)	24 (27.59)	21 (10.34)
Cameroon	1 (0.28)	0 (0.00)	0 (0.00)
EAI	0 (0.00)	0 (0.00)	1 (0.49)
Haarlem	98 (27.22)	19 (21.84)	47 (23.15)
LAM	162 (45.00)	26 (29.89)	74 (36.45)
Manu	0 (0.00)	1 (1.15)	2 (0.99)
S	11 (3.06)	2 (2.30)	5 (2.46)
T	56 (15.56)	2 (2.30)	21 (10.34)
X	13 (3.61)	4 (4.60)	15 (7.39)
Unknown	18 (5.00)	9 (10.34)	17 (8.37)

*P-value<0.000001

Peru

Lineage	Pan-susceptible (%)	MDR (%)
AFRI	22 (0.97)	0 (0.00)
BOV	6 (0.27)	0 (0.00)
Beijing	223 (9.86)	32 (9.01)
EAI	3 (0.13)	0 (0.00)
Haarlem	738 (32.63)	54 (15.21)
LAM	511 (22.59)	160 (45.07)
Manu	1 (0.04)	0 (0.00)
S	33 (1.46)	1 (0.28)
T	386 (17.06)	71 (20.00)
Ural	2 (0.09)	2 (0.56)
X	143 (6.32)	13 (3.66)
Unknown	194 (8.58)	22 (6.20)

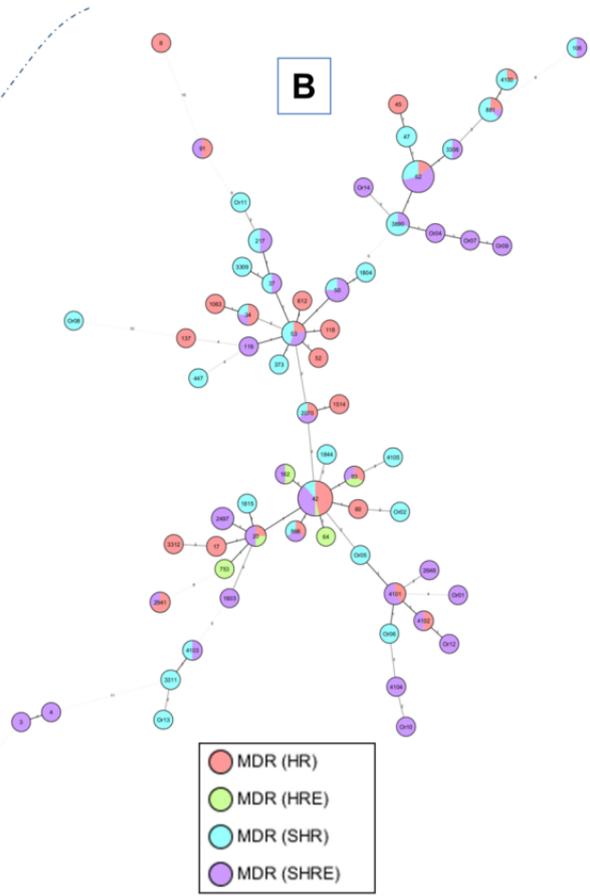
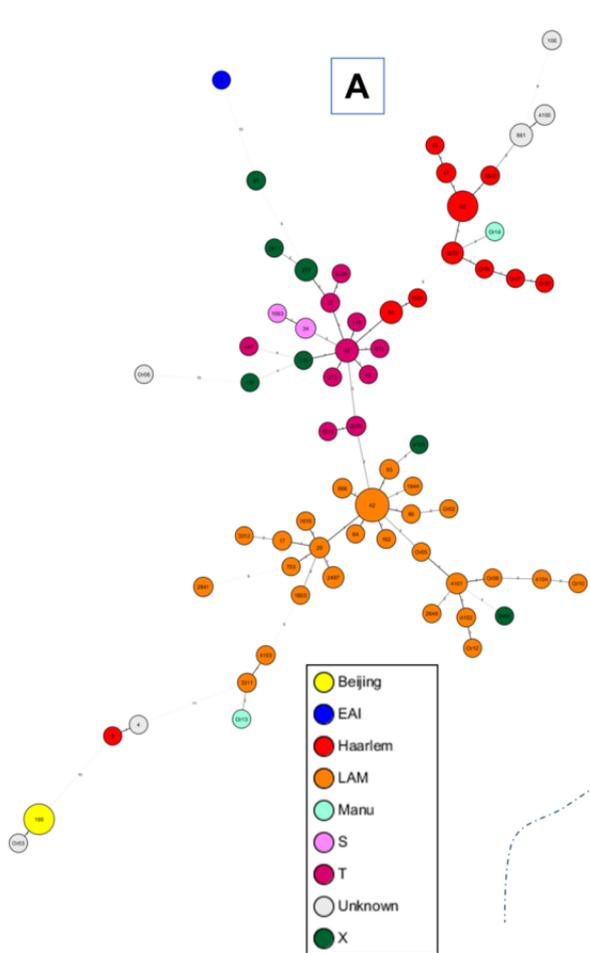
*P-value<0.000001

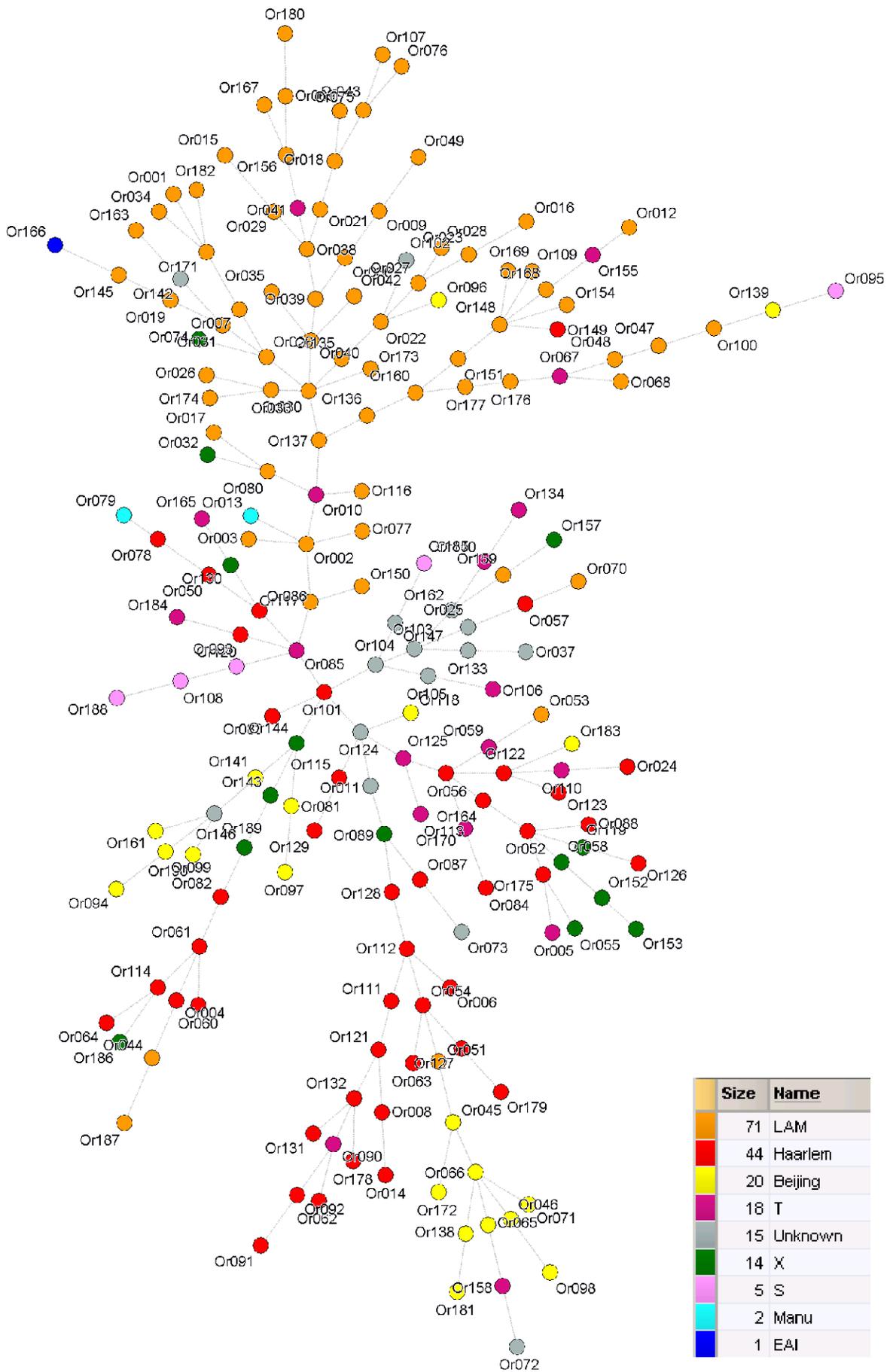
Brazil

Lineage	Pan-susceptible (%)	MDR (%)
BOV	1 (0.10)	0 (0.00)
Beijing	1 (0.10)	2 (0.68)
CAS	4 (0.40)	0 (0.00)
Cameroon	1 (0.10)	0 (0.00)
EAI	64 (6.44)	17 (5.74)
Haarlem	131 (13.18)	30 (10.14)
LAM	439 (44.16)	116 (39.19)
Manu	12 (1.21)	4 (1.35)
S	13 (1.31)	5 (1.69)
T	191 (19.22)	75 (25.34)
Unknown	91 (9.15)	16 (5.41)
Ural	5 (0.50)	0 (0.00)
X	41 (4.12)	31 (10.47)

*P-value<0.001







° Clustered strains correspond to a similar spoligotype pattern shared by 2 or more strains “within this study”; as opposed to unique strains harboring a spoligotype pattern that does not match with another strain from this study. Unique strains matching a preexisting pattern in the SITVIT database are classified as SITs, whereas in case of no match, they are designated as “orphan”.

Table 3. Distribution of MTBC lineages in function of several parameters of the study.

Lineage	Gender and Age			City of isolation				First line multi-drug resistance			
	Female	Male	Mean Age (years)	Buen-aventura	Cali	Medellin	Other cities	MDR (HR)	MDR (HRE)	MDR (SHR)	MDR (SHRE)
Beijing	7	14	37.94	16	4	0	1	0	0	1	20
EAI	0	1	56.00	0	0	0	1	1	0	0	0
Haarlem	16	31	40.20	5	2	22	18	4	0	18	25
LAM	27	47	40.84	4	6	20	44	24	6	12	32
Manu	0	2	38.50	0	0	0	2	0	0	1	1
S	1	4	43.00	1	0	0	4	3	0	1	1
T	9	12	37.75	2	2	6	11	7	0	9	5
X	6	9	35.29	1	1	9	4	2	0	6	7
Unknown	8	9	41.13	0	3	3	11	3	0	10	4